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Journal article

Between-meal sucrose-sweetened beverage consumption impairs glycaemia and lipid metabolism during prolonged sitting : A randomized controlled trial

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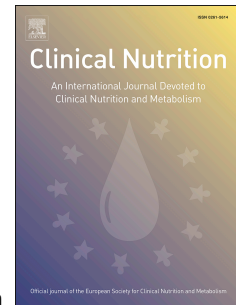
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Between-meal sucrose-sweetened beverage consumption impairs glycemia and lipid metabolism during prolonged sitting: a randomized controlled trial

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Keywords: sugar-sweetened beverages, glucose, lipids, overweight, obesity, cardiometabolic disease.

1 ABSTRACT

2 **Background & Aims:** Chronic overconsumption of sugar-sweetened beverages
3 (SSBs) is associated with unfavourable health effects, including promotion of obesity.
4 However, the acute effects of consuming SSBs on glucose and lipid metabolism
5 remain to be characterized in a real-world, post-prandial context of prolonged
6 sitting. We quantified the acute effects of between-meal SSB consumption compared
7 with water, on glucose and lipid metabolism in habitual soft drink consumers during
8 prolonged sitting.

9 **Methods:** Twenty-eight overweight or obese young adults [15 males; 23 ± 3 (mean \pm
10 SD) years, body mass index (BMI) 31.0 ± 3.6 kg/m²) participated. During
11 uninterrupted sitting and following standardized breakfast and lunch meals, each
12 participant completed two 7-hour conditions on separate days in a randomized,
13 crossover design study. For each condition, participants consumed either a sucrose
14 SSB or water mid-morning and mid-afternoon. Peak responses and total area under
15 the curve (tAUC) over 7 h for blood glucose, insulin, C-peptide, triglyceride and non-
16 esterified fatty acid (NEFA) concentrations were quantified and compared.

17 **Results:** Compared to water, SSB consumption significantly increased the peak
18 responses for blood glucose (20 ± 4 % (mean \pm SEM)), insulin (43 ± 15 %) and C-
19 peptide (21 ± 6 %) concentrations. The tAUC for all these parameters was also
20 increased by SSB consumption. The tAUC for triglycerides was 15 ± 5 % lower after
21 SSBs and this was driven by males ($P < 0.05$), as females showed no difference
22 between conditions. The tAUC for NEFAs was 13 ± 5 % lower after the SSB
23 condition ($P < 0.05$).

24 **Conclusions:** Between-meal SSB consumption significantly elevated plasma glucose
25 responses, associated with a sustained elevation in plasma insulin throughout a day of

26 prolonged sitting. The SSB-induced reduction in circulating triglycerides and NEFAs
27 indicates significant modulation of lipid metabolism, particularly in males. These
28 metabolic effects may contribute to the development of metabolic disease when SSB
29 consumption is habitual and co-occurring with prolonged sitting.

30 **Clinical Trial Registry number:** ACTRN12616000840482,
31 <https://anzctr.org.au/Trial/Registration/TrialReview.aspx?ACTRN=12616000840482>

32

33 **Abbreviations:**

34 Body Mass Index, BMI

35 Sugar-Sweetened Beverages, SSBs

36 Total Area Under the Curve, tAUC

37 Moderate-to-Vigorous intensity Physical Activity, MVPA

38 Non-esterified Fatty Acid, NEFA

39 United States, US

40 World Health Organisation, WHO

41

42 INTRODUCTION

43

44 Globally, sugar-sweetened beverages (SSBs) are the largest source of added sugars in
45 Western diets (1). SSB consumption is associated with the development of weight
46 gain, fatty liver, type 2 diabetes and cardiovascular disease (2-6). To date, most
47 studies have focused on the relationships between sugary drink consumption and
48 overweight/obesity. However, the large amount of added sugars that these drinks
49 typically contain have additional implications beyond weight control, which may
50 directly elevate risk for diabetes and cardiovascular disease (2, 3). These relate to
51 chronic post-prandial glucose excursions which contribute to pancreatic β -cell failure
52 and vascular complications as well as non-alcoholic fatty liver (7-9).

53 We have recently shown that there is significant variation across countries for
54 identically-branded soft drinks, in their total concentration of glucose and fructose, as
55 a result of global differences in primary industry sources of sugar (10). Soft drinks in
56 Australia and Europe are chiefly sweetened by sucrose (disaccharide composed of
57 50% glucose and 50% fructose), whereas formulations marketed under the same trade
58 name in the United States (US) use high-fructose corn syrup (15). It is unknown
59 whether the difference in glucose-fructose ratio between sucrose (50:50) and high-
60 fructose corn-syrup (typically 55:45) is sufficient to drive specific health effects, but
61 given the global variation in soft drink composition, there is a need to quantify the
62 magnitude by which sucrose-sweetened drinks elevate plasma glucose and insulin
63 concentrations (11, 12).

64 In addition to the adverse effects of SSBs on glycemic responses, consumption
65 in the context of prolonged uninterrupted sitting during the day would be expected to
66 exaggerate glucose and insulin excursions. Through observational and experimental

67 studies, we have shown that impaired glycemic control is an important contributor to
68 sitting-associated risk for chronic disease (13, 14). Such a perspective is important
69 given current population trends for increasingly sedentary lifestyles, as characterized
70 by time spent in prolonged sitting (15). Indeed, SSB consumption has been
71 demonstrated to co-occur with high sedentary time (13, 16-18), making this behaviour
72 a key driver of cardiometabolic risk in highly sedentary population groups (19).

73 Despite this, the acute metabolic effects of SSB consumption on both glucose
74 and lipid metabolism in a real-world context that incorporates typical daily
75 consumption levels, as well as meal patterns and prolonged sitting, have not been
76 investigated. For many young adults between-meal SSB consumption is a daily habit
77 which challenges metabolic homeostasis and potentially seeds chronic
78 cardiometabolic diseases. The purpose of this study was to quantify the acute effects
79 of between-meal sucrose-sweetened beverage consumption compared with water
80 during prolonged sitting on glucose and lipid metabolism in habitual soft drink
81 consumers.

82

83 **MATERIALS AND METHODS**

84

85 **Participants**

86 Twenty-eight inactive overweight/obese males (n=15) and females (n=13), who were
87 habitual consumers of SSBs, participated in this study. Participants were recruited via
88 posters, online advertisements, and social media. Eligibility included: age between 19
89 and 30 yr; body mass index (BMI) ≥ 25 , but ≤ 40 kg/m², SSB consumption of > 2 L
90 or more per week for at least the previous 3 months; self-reported sitting time > 5
91 h/day, and no regular moderate-to-vigorous intensity physical activity (MVPA; ≥ 150

92 min/week for > 3 months). Exclusion criteria included: being employed in a non-
93 sedentary occupation (as characterized by low demand for sitting – e.g. tradesperson),
94 currently using prescription medication that would confound interpretation of the
95 data, pregnant or currently smoking. The study was approved by the Alfred Human
96 Research Ethics Committee and all participants provided written informed consent.
97 This trial was registered with the Australian New Zealand Clinical Trials Registry at
98 <https://anzctr.org.au/Trial/Registration/TrialReview.aspx?ACTRN=12616000840482>
99 as ACTRN12616000840482.

100

101 **Study Design**

102 This randomized crossover trial was undertaken at the Baker Heart and Diabetes
103 Institute between June 2016 and August 2017. Participants completed two acute
104 single day (7 h) experimental conditions in random order with a minimum of 21 days
105 wash-out between visits. Both conditions were performed on a background of
106 uninterrupted sitting.

107 *SSB and uninterrupted sitting:* Participants sat upright in a comfortable lounge chair
108 and consumed a commercially available sucrose-sweetened beverage 90 min after a
109 standardized breakfast and lunch meal (**Fig. 1**).

110 *Water and uninterrupted sitting:* Participants sat upright in a comfortable lounge chair
111 and consumed a volume of water equal to that consumed during the SSB condition,
112 90 minutes after the breakfast and lunch meal.

113 Participants attended the laboratory on four separate occasions. During their
114 first visit, a general screening was conducted for baseline physical (height, weight,
115 waist: hip ratio, blood pressure), and biochemical (glucose, insulin, HbA1c, lipids)
116 characteristics against inclusion/exclusion criteria. During the second visit, conducted

117 seven days prior to the first experimental trial condition (visit 3), participants were
118 familiarized with each experimental condition and were instructed to complete
119 physical activity and dietary records. Participants also received (written and verbal)
120 instruction regarding the pre-experimental evening meal (including overnight fasting)
121 and physical activity restriction prior to each trial condition. Female participants were
122 asked to provide details regarding their menstrual cycle, to permit scheduling of each
123 experimental visit within the follicular phase (between days 3-10). Standardized
124 email, text message prompts and phone calls were used to maximize participant
125 compliance. To eliminate potential bias, trial condition order was randomly assigned
126 by a third party using computer-generated random numbers, stratified by sex. Study
127 personnel and participants were blinded to the condition order until the morning of the
128 first trial condition. Study investigators PV and BAK, the pathology technicians, and
129 team statisticians were blinded throughout data collection and analysis.

130

131 **Beverages**

132 The SSB was a commercially-available carbonated soft drink containing sucrose
133 (8.8g/100mL), free glucose (1.1g/100mL) and free fructose (1.1g/100mL) (10). This
134 corresponded to a total glucose (calculated final monosaccharide concentration) of
135 5.5g/100mL and a total fructose (calculated final monosaccharide concentration) of
136 5.6g/100mL(10). The two SSBs serves each provided 6% of estimated energy
137 requirements (Schofield equation, 1.5 physical activity factor) and approximated a
138 discretionary food serve (Australian dietary guidelines) (20). The total volume of soft
139 drink consumed on experimental days was reflective of levels reported among SSB
140 consumers' aged 19-30 year-olds in the recent Australian Health survey (21). The
141 average volume per serve was 376 ± 12 mL and the average total volume consumed

142 per experimental day was 752 ± 23 mL.

143 **Standardization of Diet and Physical Activity**

144 Participants were provided with a 'food pack' containing an evening meal for
145 consumption the night before each experimental day, while breakfast and lunch
146 meals were individually prepared and provided in the laboratory. Meal plans were
147 individualized to meet estimated energy requirements (Schofeld equation, 1.5
148 physical activity factor) and were based on Australian dietary intakes (21). For
149 breakfast, lunch and dinner, the macronutrient profile (as a percentage of total energy)
150 was 14-18% for protein, 48-52% for carbohydrate, and 29-32% for fat. Meals were
151 identical across conditions for each individual and provided ~33% of estimated daily
152 energy requirements. For the evening meals, participants received verbal and written
153 instructions and reminders to consume only those items within the 'food packs'. They
154 were also instructed and reminded to record their dietary intake in the provided diary
155 and to refrain from consuming alcohol and caffeine in the 24 h preceding each
156 experimental condition. Weighed/measured food records were individually completed
157 and dietary intakes assessed using Australian-specific dietary analysis software
158 (FoodWorks: Xyris Software, Version 8, AUS).

159 To minimize any potential effects of physical activity, participants were
160 instructed to avoid moderate and/or vigorous exercise for at least 48 h prior to each
161 experimental condition. To confirm this, participants kept an activity diary and wore a
162 triaxial accelerometer (GTX3+; Actigraph, Pensacola, FL) to objectively assess their
163 activity levels during waking hours for seven consecutive days before the condition
164 (defined as the habitual period) and during the experimental condition day. They
165 were instructed to wear the accelerometer on the right hip during all waking hours,
166 unless doing water-based activities. The 1-min epoch activity data (for waking hours)

167 were then processed using a cut off < 100 counts/min define sedentary time (22, 23).
168 Freedson's cut offs were used to differentiate moderate-to-vigorous-intensity activity
169 (counts/min \geq 1,952) from light-intensity activity (100 –1,951 counts/min) (24). Total
170 time was calculated as the sum of time spent in all activities (sedentary, light and
171 MVPA). Data are reported as averages for valid days (days with > 10 hours wear and
172 no minutes with counts \geq 20,000).

173

174 **Study Protocol**

175 After a minimum 10 h overnight fast, participants reported to the laboratory at 0715 h.
176 After voiding, and once anthropometric measurements were obtained, an indwelling
177 catheter was inserted into an antecubital vein and fasting blood samples were
178 collected before (-1 h) and after (0 h) a 1 h seated steady-state period.

179 At 0 h participants consumed the standardized breakfast meal with the time
180 taken to consume (< 20 min) replicated in subsequent conditions (**Fig. 1**). At 3 h
181 participants consumed lunch (< 20 min). Ninety-minutes after each meal, participants
182 consumed individualized volumes of the SSB or water within 10 min. Postprandial
183 blood samples were collected at 30-minute intervals over each 7 h experimental
184 condition. A total of 273 mL of blood was taken from each participant during an
185 experimental trial. Participants had access to internet services, standardized television
186 and DVD viewing and reading materials (newspapers and magazines) during the two
187 experimental conditions. To minimize unscheduled physical activity, standardized
188 lavatory visits were incorporated into the protocol immediately following SSB or
189 water consumption (1.5 h and 4.5 h) for each trial.

190

191

[Insert Fig. 1 here]

192

193 **Biochemical Analyses**

194 All blood for screening and the experimental conditions was collected in to
195 appropriate tubes (BD VacutainerTM, Franklin Lakes, NJ, USA) for determination of
196 concentrations of glucose, insulin, C-peptide, HbA1c, total cholesterol, HDL-
197 cholesterol, LDL-cholesterol, triglycerides, NEFAs and human chorionic
198 gonadotrophin (for females). All analyses except for NEFAs were conducted at the
199 Alfred Hospital, Department of Pathology according to clinical diagnostic standards
200 (National Association of Testing Authorities accredited). Plasma glucose was
201 measured using the hexokinase method. Serum insulin and C-peptide were measured
202 using a chemiluminescent microparticle immunoassay (Architect ci16200; Abbott
203 Diagnostics, Santa Clara, CA). At visits 3 and 4, baseline and hourly samples were
204 drawn into EDTA tubes, centrifuged ($2000 \times g$ for 15min at 4°C) and the plasma
205 stored at -80°C for later analysis of NEFAs using a commercially available kit (Waco
206 Diagnostics, Richmond, VA, USA). Insulin resistance was estimated from fasting
207 glucose by using a computer-based homeostasis model assessment system (HOMA2-
208 IR) provided by the Oxford Centre for Diabetes, Endocrinology, and Metabolism
209 ([http:// www.dtu.ox.ac.uk/homa](http://www.dtu.ox.ac.uk/homa)). Similar previous studies have used and validated
210 this approach (25).

211

212 **Statistical Analyses**

213 Study data were collated and managed using REDCap electronic data capture tools
214 hosted at [Baker Heart and Diabetes Institute] (26). Physical characteristics were
215 compared between males and females using an unpaired two-tailed Student's t test.
216 Anthropometric, dietary, and accelerometer-derived physical activity data before each

217 of the respective trial conditions are presented in **Supplementary Table 1**. The small
218 but statistically significant difference in sedentary time (48 h prior to experimental
219 visits) had no effect on endpoint analyses and was therefore not included as a
220 covariate.

221 Plasma glucose in both the morning and the afternoon was the primary
222 outcome measure, with sample size determined by power calculations based on our
223 previous studies (13, 14). To allow examination of differential effects of the
224 interventions throughout the day, the study was powered at a β value of 80% to detect
225 a 15% minimum difference in glycaemia (based on a standard deviation of the
226 difference of 25%) after both the morning and the afternoon drink at an alpha level of
227 0.025 (to accommodate dual endpoints). Peak plasma glucose in response to each
228 drink in the morning [(1.5 – 3 h (Drink 1))] and afternoon [(4.5 -6 h (Drink 2))] was
229 calculated. Total area under the curve (AUC) (trapezoidal method using a baseline of
230 zero) over the 7 h intervention was also calculated for glucose, insulin, C-peptide,
231 triglycerides and NEFAs. Generalized linear mixed models (with random intercepts)
232 were used to evaluate the differential effects of the experimental conditions on the
233 selected outcomes using Stata 14 (StataCorp LP, College Station, Texas, USA). All
234 models were adjusted for potential covariates explaining residual outcome variance
235 (age, sex, and BMI), baseline values, and period effects (treatment order). Residuals
236 were examined for serial correlation, heteroscedasticity and normality. Substantial
237 departures from model assumptions were not observed. Sex-by-condition, interactions
238 were performed for each tAUC outcome measure. Statistical significance was set at P
239 < 0.05 . Data are expressed as mean \pm SEM unless otherwise stated.

240

241 **RESULTS**

242

243 **Participant Characteristics**

244 Thirty-three participants were randomized and familiarized, but five withdrew prior to
245 the first experimental condition (**Supplementary Fig. 1**). As such, twenty-eight
246 participants [15 males, 13 females; 23 ± 3 years, BMI 31.0 ± 3.6 kg/m²; (mean \pm SD)]
247 commenced and completed all trial conditions (**Table 1**). There were no significant
248 differences in baseline variables between sexes except for HDL-cholesterol which
249 was higher in women.

250

251

[Insert Table 1 here]

252

253 **Glycemic Responses**

254 The average volume of the SSB and water, and average amount of sugars (sucrose,
255 glucose, fructose, total glucose and total fructose, calculated final monosaccharide
256 concentration) consumed during the trial conditions are presented in **Supplementary**
257 **Table 2**. For the SSB intervention, the average amount of total glucose (calculated
258 final monosaccharide) was 20.7 ± 0.6 g per serve and 41.4 ± 1.3 g per trial day and
259 total fructose (calculated final monosaccharide) was 21.1 ± 0.7 g per serve and $42.1 \pm$
260 1.3 g per trial day.

261

262

263

264

265

266

Fig. 2 (A-C) shows the plasma glucose, insulin and C-peptide concentrations during each of the experimental conditions. Compared to water, between-meal SSB consumption significantly increased peak plasma glucose, insulin and C-peptide concentrations both in the morning by 20 ± 4 % (mean \pm SEM), 43 ± 15 % and 21 ± 6 %, and in the afternoon by 8 ± 3 %, 35 ± 14 % and 15 ± 6 %; all $P < 0.05$ (**Fig. 2; D-F**). The tAUCs were significantly higher for the SSB intervention compared to water

267 for glucose, insulin and C-peptide by 5 ± 1 %, 26 ± 9 % and 11 ± 3 %, respectively;
268 all $P < 0.05$.

269

270 **[Insert Fig. 2 here]**

271

272 **Lipid Responses**

273 **Fig. 3** shows the plasma triglyceride concentrations during each of the
274 experimental conditions for males and females. The tAUC for plasma triglycerides
275 was significantly lower after SSB consumption compared to water by 15 ± 5 % ($P <$
276 0.05). The reduction for the morning period was 13 ± 4 %) and the afternoon was 18
277 ± 6 % ($P < 0.05$ for both). There was a significant sex-by-condition interaction effect
278 for the triglyceride tAUC which corresponded to a 24 ± 5 % reduction in males after
279 the SSB compared to the water condition ($P < 0.05$). Females had significantly lower
280 triglyceride levels than males at baseline but showed no difference between
281 conditions (**Fig. 3**).

282

283 **[Insert Fig. 3 here]**

284

285 **Fig. 4** shows the NEFA concentrations during each of the experimental
286 conditions. There was a trend for higher baseline NEFA concentration in the SSB
287 trial, but after adjustment (see Statistical Analysis), the tAUC for NEFA
288 concentrations was significantly lower after SSB consumption compared to water (by
289 13 ± 5 %; $P < 0.05$). This was evident in both the morning (10 ± 4 %) and in the
290 afternoon (21 ± 9 %; $P < 0.05$). There was no significant sex-by-condition interaction
291 for NEFA concentration, nor any other outcome.

292

293

[Insert Fig. 4 here]

294

295 **DISCUSSION**

296 Consumption of a sucrose-sweetened beverage in a pattern typical of habitual soft
297 drink consumers, and in the context of normal meals and prolonged sitting, elevated
298 peak plasma glucose concentration by 20% compared to water. This was associated
299 with a sustained 26% elevation in plasma insulin throughout the day. These effects
300 were observed in parallel with modulation of parameters associated with lipid
301 metabolism. Plasma triglyceride concentration was 15% lower after sucrose-
302 sweetened beverage consumption compared to water, an effect limited to men, where
303 values were reduced by 24%. In addition, NEFAs were reduced by 13% after sucrose-
304 sweetened beverage consumption compared with water. These effects are relevant to
305 typical daily consumption levels (27), are in the context of real world behaviour
306 patterns (regular meals and prolonged sitting) and are quantitated in comparison to
307 water which is considered the optimal alternative to SSB consumption in the general
308 community (28).

309 The effects of sucrose-sweetened beverage consumption on glucose and fat
310 metabolism are of interest because these formulations are higher in glucose than high-
311 fructose corn syrup formulations (10). The differential effects of glucose and fructose
312 consumption are most likely due to glucose being absorbed from the small intestine
313 into the blood where it elevates blood glucose concentration and stimulates the
314 pancreas to produce insulin(29). In contrast, fructose is primarily metabolized in the
315 liver (30), stimulating glycogenesis, gluconeogenesis and lipogenesis. Distinct from
316 glucose, fructose does not acutely increase blood glucose or insulin concentration

317 (31).

318 Specific effects of sucrose-sweetened drinks, which we have shown to be 22%
319 higher in glucose than high-fructose corn syrup sweetened drinks, may relate to
320 induction of high and variable plasma glucose and insulin levels (10). Chronic post-
321 prandial glucose excursions and variability contribute to pancreatic β -cell failure and
322 progression to late-stage diabetes (32).

323

324 *Glucose metabolism*

325 The greatest difference in post drink plasma glucose in the current study was observed
326 after the first drink. Moderation of the increase in plasma glucose after the second
327 drink was achieved through elevated insulin levels established after the first drink and
328 sustained throughout the day (second meal effect) (6, 9, 33, 34).

329 Previous research has focused predominantly on the relationship between
330 SSBs and weight gain (35-37). Some studies examining physiological responses to
331 sugar consumption have examined single doses of individual sugars (e.g. sucrose,
332 glucose or fructose) on a fasting background and over relatively short follow up
333 periods of two hours or less (12, 38, 39). These studies demonstrate large excursions
334 in blood glucose (up to 60%) and insulin in response to consumption of glucose
335 drinks, but are less relevant to the real-world scenario of mixed sugar consumption
336 associated with SSBs in the context of meals (11, 29, 39, 40). Other studies have
337 considered SSB consumption in the context of meals, but have examined very high
338 SSB ‘doses’ supplying 25% of daily energy requirements (39).

339 The SSB “dose” delivered in our study (12% of daily energy requirements,
340 **Supplementary Table 2**) is highly relevant to current global consumption trends. In
341 the age group corresponding to the current study (19 to 30 years), where SSB

342 consumption is greatest, the top 10% highest consumers in the Australian Health
343 Survey drank more than 1 L of SSBs, peaking at 1.5 L (28 teaspoons or 110 g) for
344 males on the day prior to interview (21). Alarming, these consumption levels far
345 exceed the current World Health Organisation (WHO) recommendations to limit
346 intake of total sugars to less than 50 g (approximately 12 teaspoons) per day (41).

347 Compounding the negative health impact of sugar over-consumption are
348 concurrent population trends for low levels of physical activity and prolonged periods
349 of sedentary time that are characterized by the absence of skeletal muscle contractile
350 activity (16). Despite strong evidence indicating that exercise can mitigate some of
351 the detrimental effects of high sugar intake, independently of energy balance (42, 43),
352 recent estimates suggest that sitting occupies the majority of the waking hours in
353 adults (between 7 and 10 hours per day) (18). Consistent with these findings, our
354 study participants spent approximately 10 hours per day sedentary (**Supplementary**
355 **Table 2**). We have established through recent observational and experimental studies
356 that impaired glycemic control is an important contributor to sitting-associated risk
357 (13, 14). The current findings are thus highly relevant in terms of characterizing the
358 metabolic impact of SSB consumption against a background of high levels of daily
359 sitting time.

360 Our results contrast with a previous study employing a very similar protocol
361 and SSB consumption pattern (two 355mL sucrose-sweetened beverages; 75 g of
362 sucrose), but examining interstitial glucose rather than plasma glucose (44). The
363 finding that sucrose-sweetened beverage consumption in this previous study did not
364 affect interstitial glucose compared with water brings into question the reliability of
365 interstitial glucose measures to monitor acute glucose changes (45, 46). The current
366 study clearly demonstrates substantial effects of sucrose-sweetened beverage

367 consumption on both plasma glucose and insulin which are likely to be detrimental in
368 regular consumers of sucrose-sweetened soft drinks and particularly in those with
369 elevated cardiometabolic risk factors.

370

371 *Lipid metabolism*

372 Glucose-induced insulin elevation also has consequences for fat metabolism, as a
373 result of insulin-mediated suppression of liver triglyceride production and lipolysis in
374 favour of glucose catabolism (38). Through this mechanism, excess glucose
375 consumption may contribute to liver fat accumulation. We observed a large SSB-
376 induced elevation in insulin, which may further exacerbate the detrimental effects of
377 fructose on lipid metabolism and liver fat accumulation (47). The elevation in insulin
378 was associated with lower plasma triglycerides, an effect driven entirely by the
379 response seen in males and consistent with suppressed liver triglyceride production in
380 very low-density lipoprotein (VLDL). These data align with known sex differences in
381 liver insulin sensitivity, in that obese men are more sensitive to glucose and insulin
382 induced suppression of liver VLDL-triglyceride production than obese women (48).
383 Alternatively, other potential explanations include that reduced adipose tissue
384 lipolysis and NEFA flux could also contribute to reduced hepatic VLDL production
385 (11). These possibilities suggest that sucrose-sweetened beverage consumption may
386 predispose men to an elevated risk for fatty liver disease. This is particularly
387 concerning given that young males lead SSB consumption in terms of both population
388 prevalence and volumes consumed (21). In addition to the gender-specific effects on
389 plasma triglycerides, NEFAs were also reduced by sucrose-sweetened beverage
390 consumption suggesting suppression of lipolysis in both men and women.(11)

391

392 *Strengths and limitations*

393 This was an appropriately powered, controlled randomized cross-over study
394 incorporating young adult male and female participants, who were typical consumers
395 of SSBs . Participants were their own controls, enhancing both the internal validity
396 and reliability of our data, and demonstrated good compliance with consumption of
397 all standardized meals and beverages during lead-in periods. Additionally, there was
398 stringent control of potential confounding variables such as diet, sedentary behaviour
399 and physical activity, through use of weighed food records and objectively measured
400 sedentary and physical activity behaviours. Nevertheless, in interpreting these
401 findings it is important to consider some limitations that future studies could address.
402 First, due to the acute nature of this study we cannot speculate on the possible
403 longer-term effects of sustained sucrose-sweetened beverage consumption. Second,
404 blinding of research participants to experimental conditions (water, SSB) was not
405 possible due to the nature of the intervention. Third, our sex-specific analysis was
406 exploratory, however, the results suggest that future research on these differences is
407 warranted. Finally, the acute effects observed in habitual SSB consumers cannot be
408 generalized amongst other populations including the non-obese, children/adolescents
409 (< 19 years), and middle aged/older adults (> 31 years).

410

411 **CONCLUSION**

412 Compared with water, consumption of sucrose-sweetened beverages significantly
413 elevates post-drink plasma glucose in association with a sustained elevation in plasma
414 insulin throughout a day of prolonged sitting. The SSB-induced reduction in
415 circulating triglycerides and NEFAs indicates significant suppression of lipid
416 metabolism, particularly in males. These metabolic effects may contribute to the

417 development of metabolic disease when SSB consumption in the context of prolonged
418 sitting is habitual.

419

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425

426 **Statement of Authorship**

427 The authors' responsibilities were as follows—BAK, NO, DWD, RNL, JH, BLD,
428 MG, GLJ, PCD, and MFF: designed the research; PV, MFF, and MRL: conducted
429 the research; PV, PCD and BAK, analyzed data or performed statistical analysis;
430 GLW and NDC provided clinical support during data collection; PV, PCD, NO,
431 DWD, RNL, JH, GLJ and BAK: wrote and participated in critical revision of the
432 manuscript for intellectual content.; and PV: had primary responsibility for final
433 content of the manuscript. None of the authors had any conflicts of interest regarding
434 this manuscript.

435

436 **Conflict of Interest**

437 The authors declare that they have no conflict of interest.

438

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References

1. Singh GM, Micha R, Khatibzadeh S, Shi P, Lim S, Andrews KG, et al. Global, Regional, and National Consumption of Sugar-Sweetened Beverages, Fruit Juices, and Milk: A Systematic Assessment of Beverage Intake in 187 Countries. *PLoS One*. 2015;10(8):e0124845.
2. Malik VS, Pan A, Willett WC, Hu FB. Sugar-sweetened beverages and weight gain in children and adults: a systematic review and meta-analysis. *Am J Clin Nutr*. 2013;98(4):1084-102.
3. Vartanian LR, Schwartz MB, Brownell KD. Effects of soft drink consumption on nutrition and health: a systematic review and meta-analysis. *Am J Public Health*. 2007;97(4):667-75.
4. Imamura F, O'Connor L, Ye Z, Mursu J, Hayashino Y, Bhupathiraju SN, et al. Consumption of sugar sweetened beverages, artificially sweetened beverages, and fruit juice and incidence of type 2 diabetes: systematic review, meta-analysis, and estimation of population attributable fraction. *BMJ*. 2015;351:h3576.
5. Malik VS, Popkin BM, Bray GA, Despres JP, Hu FB. Sugar-sweetened beverages, obesity, type 2 diabetes mellitus, and cardiovascular disease risk. *Circulation*. 2010;121(11):1356-64.
6. Ma J, Fox CS, Jacques PF, Speliotes EK, Hoffmann U, Smith CE, et al. Sugar-sweetened beverage, diet soda, and fatty liver disease in the Framingham Heart Study cohorts. *J Hepatol*. 2015;63(2):462-9.
7. Russell ND, Cooper ME. 50 years forward: mechanisms of hyperglycaemia-driven diabetic complications. *Diabetologia*. 2015;58(8):1708-14.
8. Maersk M, Belza A, Stodkilde-Jorgensen H, Ringgaard S, Chabanova E, Thomsen H, et al. Sucrose-sweetened beverages increase fat storage in the liver, muscle, and visceral fat depot: a 6-mo randomized intervention study. *Am J Clin Nutr*. 2012;95(2):283-9.
9. Mah E, Bruno RS. Postprandial hyperglycemia on vascular endothelial function: mechanisms and consequences. *Nutr Res*. 2012;32(10):727-40.
10. Varsamis P, Larsen RN, Dunstan DW, Jennings GL, Owen N, Kingwell BA. The sugar content of soft drinks in Australia, Europe and the United States. *Med J Aust*. 2017;206(10):454-5.
11. Chong MF, Fielding BA, Frayn KN. Mechanisms for the acute effect of fructose on postprandial lipemia. *Am J Clin Nutr*. 2007;85(6):1511-20.
12. Jameel F, Phang M, Wood LG, Garg ML. Acute effects of feeding fructose, glucose and sucrose on blood lipid levels and systemic inflammation. *Lipids Health Dis*. 2014;13:195.
13. Dunstan DW, Kingwell BA, Larsen R, Healy GN, Cerin E, Hamilton MT, et al. Breaking up prolonged sitting reduces postprandial glucose and insulin responses. *Diabetes Care*. 2012;35(5):976-83.
14. Dempsey PC, Larsen RN, Sethi P, Sacre JW, Straznicky NE, Cohen ND, et al. Benefits for Type 2 Diabetes of Interrupting Prolonged Sitting With Brief Bouts of Light Walking or Simple Resistance Activities. *Diabetes Care*. 2016;39(6):964-72.
15. Australian Bureau of Statistics. Australian Health Survey: Physical Activity, 2011-12 2011-12 [cited 2016 20/01/16]. Available from:

<http://www.abs.gov.au/ausstats/abs@.nsf/Lookup/4364.0.55.004Chapter1002011-12>.

16. Fletcher EA, McNaughton SA, Crawford D, Cleland V, Della Gatta J, Hatt J, et al. Associations between sedentary behaviours and dietary intakes among adolescents. *Public Health Nutr.* 2018;21(6):1115-22.
17. Chau JY, Grunseit AC, Chey T, Stamatakis E, Brown WJ, Matthews CE, et al. Daily sitting time and all-cause mortality: a meta-analysis. *PLoS One.* 2013;8(11):e80000.
18. Healy GN, Wijndaele K, Dunstan DW, Shaw JE, Salmon J, Zimmet PZ, et al. Objectively measured sedentary time, physical activity, and metabolic risk: the Australian Diabetes, Obesity and Lifestyle Study (AusDiab). *Diabetes Care.* 2008;31(2):369-71.
19. Rendell MS, Jovanovic L. Targeting postprandial hyperglycemia. *Metabolism.* 2006;55(9):1263-81.
20. Schofield WN. Predicting basal metabolic rate, new standards and review of previous work. *Hum Nutr Clin Nutr.* 1985;39 Suppl 1:5-41.
21. Australian Bureau of Statistics. Australian Health Survey 2011-2012: Nutrition First Results - Foods and Nutrients: Consumption of Sweetened Beverages. 2015 [Available from: <http://www.abs.gov.au/ausstats/abs@.nsf/Lookup/4364.0.55.007main+features7102011-12>].
22. Ekelund U, Griffin SJ, Wareham NJ. Physical activity and metabolic risk in individuals with a family history of type 2 diabetes. *Diabetes Care.* 2007;30(2):337-42.
23. Healy GN, Dunstan DW, Salmon J, Cerin E, Shaw JE, Zimmet PZ, et al. Objectively measured light-intensity physical activity is independently associated with 2-h plasma glucose. *Diabetes Care.* 2007;30(6):1384-9.
24. Freedson PS, Melanson E, Sirard J. Calibration of the Computer Science and Applications, Inc. accelerometer. *Med Sci Sports Exerc.* 1998;30(5):777-81.
25. Wallace TM, Levy JC, Matthews DR. Use and abuse of HOMA modeling. *Diabetes Care.* 2004;27(6):1487-95.
26. Harris PA, Taylor R, Thielke R, Payne J, Gonzalez N, Conde JG. Research electronic data capture (REDCap)--a metadata-driven methodology and workflow process for providing translational research informatics support. *Journal of biomedical informatics.* 2009;42(2):377-81.
27. Australian Bureau of Statistics. Australian Health Survey: Consumption of Added Sugars 2016 [10/05/2016]. Available from: <http://www.abs.gov.au/AUSSTATS/abs@.nsf/DetailsPage/4364.0.55.0112011-12?OpenDocument>.
28. Johnson RK, Lichtenstein AH, Anderson CAM, Carson JA, Després JP, Hu FB, et al. Low-Calorie Sweetened Beverages and Cardiometabolic Health: A Science Advisory From the American Heart Association. *Circ J.* 2018;138:1-15.
29. Teff KL, Elliott SS, Tschöp M, Kieffer TJ, Rader D, Heiman M, et al. Dietary fructose reduces circulating insulin and leptin, attenuates postprandial suppression of ghrelin, and increases triglycerides in women. *J Clin Endocrinol Metab.* 2004;89(6):2963-72.
30. Jang C, Hui S, Lu W, Cowan AJ, Morscher RJ, Lee G, et al. The Small Intestine Converts Dietary Fructose into Glucose and Organic Acids. *Cell Metab.* 2018;27(2):351-61.e3.

31. Bray GA, Nielsen SJ, Popkin BM. Consumption of high-fructose corn syrup in beverages may play a role in the epidemic of obesity. *Am J Clin Nutr.* 2004;79(4):537-43.
32. Kaiser N, Leibowitz G, Neshier R. Glucotoxicity and beta-cell failure in type 2 diabetes mellitus. *J Pediatr Endocrinol Metab.* 2003;16(1):5-22.
33. Maersk M, Belza A, Holst JJ, Fenger-Gron M, Pedersen SB, Astrup A, et al. Satiety scores and satiety hormone response after sucrose-sweetened soft drink compared with isocaloric semi-skimmed milk and with non-caloric soft drink: a controlled trial. *Eur J Clin Nutr.* 2012;66(4):523-9.
34. Loader J, Meziat C, Watts R, Lorenzen C, Sigaucho-Roussel D, Stewart S, et al. Effects of Sugar-Sweetened Beverage Consumption on Microvascular and Macrovascular Function in a Healthy Population. *Arterioscler Thromb Vasc Biol.* 2017;37(6):1250-60.
35. Pereira MA. Sugar-sweetened and artificially-sweetened beverages in relation to obesity risk. *Adv Nutr.* 2014;5(6):797-808.
36. Gibson S. Sugar-sweetened soft drinks and obesity: a systematic review of the evidence from observational studies and interventions. *Nutr Res Rev.* 2008;21(2):134-47.
37. Forshee RA, Anderson PA, Storey ML. Sugar-sweetened beverages and body mass index in children and adolescents: a meta-analysis. *Am J Clin Nutr.* 2008;87(6):1662-71.
38. Stanhope KL, Schwarz JM, Keim NL, Griffen SC, Bremer AA, Graham JL, et al. Consuming fructose-sweetened, not glucose-sweetened, beverages increases visceral adiposity and lipids and decreases insulin sensitivity in overweight/obese humans. *J Clin Invest.* 2009;119(5):1322-34.
39. Stanhope KL, Griffen SC, Bair BR, Swarbrick MM, Keim NL, Havel PJ. Twenty-four-hour endocrine and metabolic profiles following consumption of high-fructose corn syrup-, sucrose-, fructose-, and glucose-sweetened beverages with meals. *Am J Clin Nutr.* 2008;87(5):1194-203.
40. Stanhope KL, Griffen SC, Bremer AA, Vink RG, Schaefer EJ, Nakajima K, et al. Metabolic responses to prolonged consumption of glucose- and fructose-sweetened beverages are not associated with postprandial or 24-h glucose and insulin excursions. *Am J Clin Nutr.* 2011;94(1):112-9.
41. World Health Organization. Sugars intake for adults and children. Geneva; 2015.
42. Egli L, Lecoultre V, Theytaz F, Campos V, Hodson L, Schneiter P, et al. Exercise prevents fructose-induced hypertriglyceridemia in healthy young subjects. *Diabetes.* 2013;62(7):2259-65.
43. Varsamis P, Walther G, Share B, Taylor F, Stewart S, Lorenzen C, et al. Transient endothelial dysfunction induced by sugar-sweetened beverage consumption may be attenuated by a single bout of aerobic exercise. *Microvasc Res.* 2018;115:8-11.
44. Manders RJ, Pennings B, Beckers CP, Aipassa TI, van Loon LJ. Prevalence of daily hyperglycemia in obese type 2 diabetic men compared with that in lean and obese normoglycemic men: effect of consumption of a sucrose-containing beverage. *Am J Clin Nutr.* 2009;90(3):511-8.
45. Kulcu E, Tamada JA, Reach G, Potts RO, Lesho MJ. Physiological differences between interstitial glucose and blood glucose measured in human subjects. *Diabetes Care.* 2003;26(8):2405-9.

46. Boyne MS, Silver DM, Kaplan J, Saudek CD. Timing of changes in interstitial and venous blood glucose measured with a continuous subcutaneous glucose sensor. *Diabetes*. 2003;52(11):2790-4.
47. Schwarz JM, Noworolski SM, Erkin-Cakmak A, Korn NJ, Wen MJ, Tai VW, et al. Effects of Dietary Fructose Restriction on Liver Fat, De Novo Lipogenesis, and Insulin Kinetics in Children With Obesity. *Gastroenterology*. 2017;153(3):743-52.
48. Mittendorfer B, Yoshino M, Patterson BW, Klein S. VLDL Triglyceride Kinetics in Lean, Overweight, and Obese Men and Women. *J Clin Endocrinol Metab*. 2016;101(11):4151-60.

Tables

Table 1.

Participant Characteristics

Characteristic	Total population	Males ¹	Females	P Value ²
Sex, <i>n</i> (%)		15 (54)	13 (46)	0.411
Age, <i>y</i>	23 ± 3	23 ± 3	24 ± 3	0.226
BMI, <i>kg/m</i> ²	31.0 ± 3.6	30.7 ± 3.1	31.3 ± 4.2	0.678
Waist circumference, <i>cm</i>	98.2 ± 13.4	102.3 ± 11.5	93.5 ± 14.3	0.084
HbA _{1c} , %	5.3 ± 0.3	5.3 ± 0.3	5.3 ± 0.2	0.881
HbA _{1c} , <i>mmol/mol</i>	34.8 ± 3.0	34.9 ± 3.8	34.8 ± 1.9	0.934
Fasting glucose, <i>mmol/L</i>	4.9 ± 0.4	5.0 ± 0.4	4.8 ± 0.4	0.750
Fasting insulin, <i>μU/mL</i>	13.7 ± 6.0	13.7 ± 5.8	13.6 ± 6.4	0.993
Fasting cholesterol, <i>mmol/L</i>				
Total	4.6 ± 1.1	4.7 ± 1.1	4.4 ± 1.1	0.537
LDL	2.8 ± 0.9	3.0 ± 0.9	2.7 ± 0.9	0.347
HDL	1.2 ± 0.3	1.0 ± 0.2	1.3 ± 0.4	0.004
Fasting triglycerides, <i>mmol/L</i>	1.2 ± 0.7	1.5 ± 0.9	1.0 ± 0.4	0.067
HOMA2%B	86 ± 42	92 ± 51	80 ± 28	0.431
HOMA2%S	135 ± 45	125 ± 45	147 ± 42	0.205
HOMA2-IR value, <i>AU</i>	0.9 ± 0.7	1.0 ± 0.8	0.8 ± 0.5	0.356
Systolic blood pressure, <i>mmHg</i>	112 ± 12	115 ± 12	109 ± 10	0.122
Diastolic blood pressure, <i>mmHg</i>	70 ± 8	68 ± 7	73 ± 8	0.102
Heart rate, <i>bpm</i>	73 ± 10	70 ± 9	77 ± 11	0.089

Abbreviations: AU, arbitrary units; BMI, body mass index; HDL, high-density lipoprotein; HOMA2%B homeostasis model assessment of estimated beta cell function; HOMA2%S, homeostasis model assessment of insulin sensitivity; HOMA2-IR homeostasis model assessment of insulin resistance index; LDL, low-density lipoprotein.

¹ Data are mean ± SD or number (%).

² Males and females compared with unpaired two-tailed Student's t test (continuous variables) and Chi-square test (categorical variables).

Figures

Fig. 1

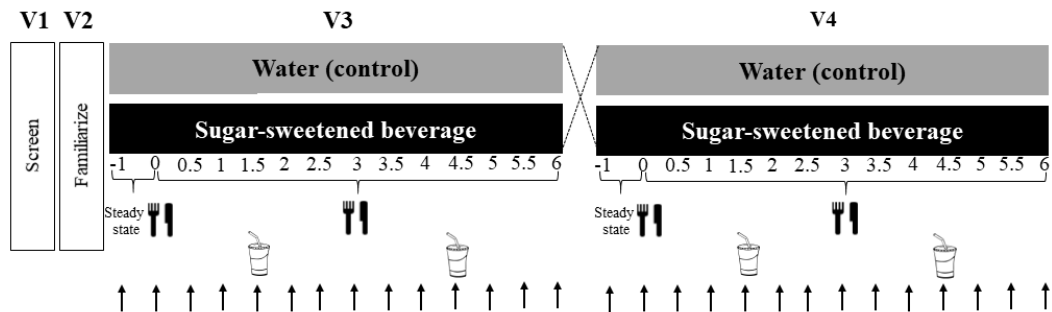


Fig. 1 Experimental randomized, cross-over study design and study day protocol for each condition with measurement time-points (in hours). Participants visited the laboratory on four separate occasions. The two trial conditions (visits 3 & 4) were completed in a randomized order separated by a minimum 21-day washout. All participants consumed standardized breakfast and lunchtime meals (🍴) at 0 h and 3 h. At 1.5 h and 4.5 h, a water or a SSB (🥤) was consumed. Blood (↑) was collected half hourly for glucose, insulin and C-peptide and hourly for triglycerides and non-esterified fatty acids (NEFAs).

Fig. 2

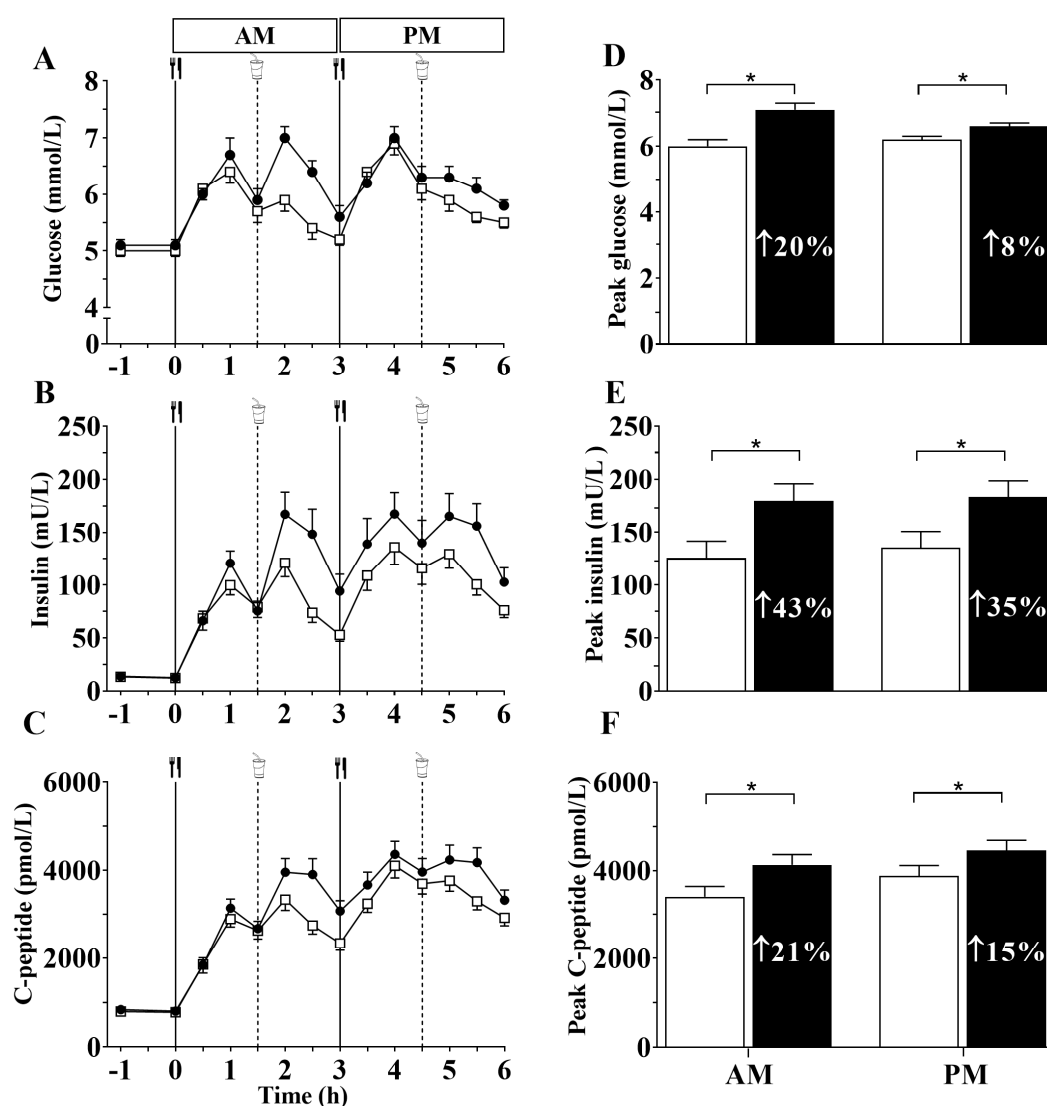


Fig. 2 Fasting (-1 and 0 h) and postprandial plasma glucose (A), serum insulin (B) and serum C-peptide (C) concentrations measured during water (open squares) and SSB conditions (closed circles). Solid vertical lines indicate timing the breakfast (0 h) and lunch (3.0 h) meals. Vertical dashed lines indicate the timing of drink 1 (1.5 h) and drink 2 (4.5 h). Peak drink responses in the morning (AM; 1.5-3 h) and afternoon (PM; 4.5-6 h) for plasma glucose (D), serum insulin (E) and serum C-peptide (F) concentrations measured during water (white bars) and SSB conditions

(black bars). Values within the bars indicate the percentage change compared to the water condition. All data are presented as mean \pm SEM. * Difference between water and SSB condition ($P < 0.05$).

ACCEPTED MANUSCRIPT

Fig.3

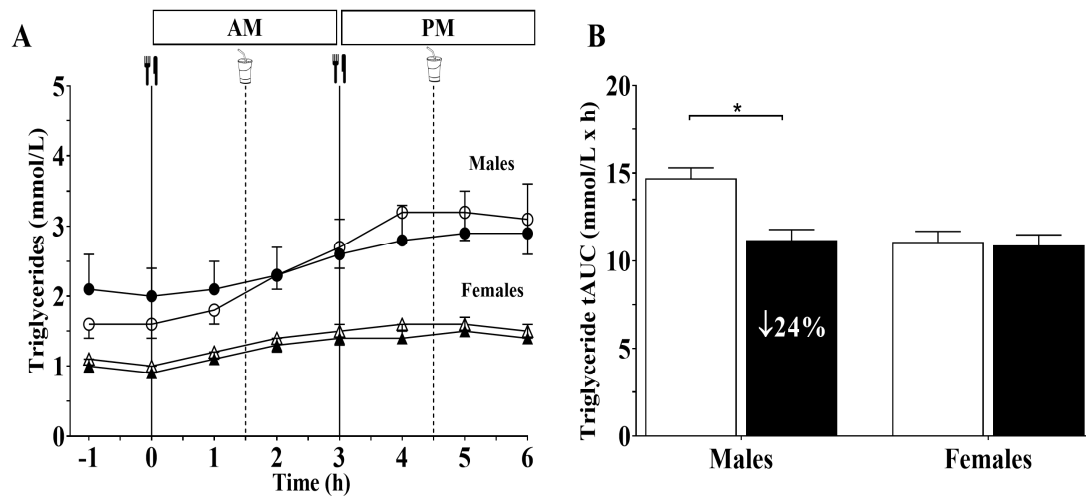


Fig. 3 Fasting and postprandial plasma triglyceride concentrations measured during water (open circle and open triangle) and SSB (closed circle and closed triangle) conditions for males (n=15) (circles) and females (n=13) (triangles) (A). Triglyceride total area under the curves (tAUC) per trial condition for males and females (B) [Water (white bars) and SSB (black bars) (B)]. All data are presented as mean \pm SEM. * Sex-by-condition interaction effect ($P < 0.05$).

Fig.4

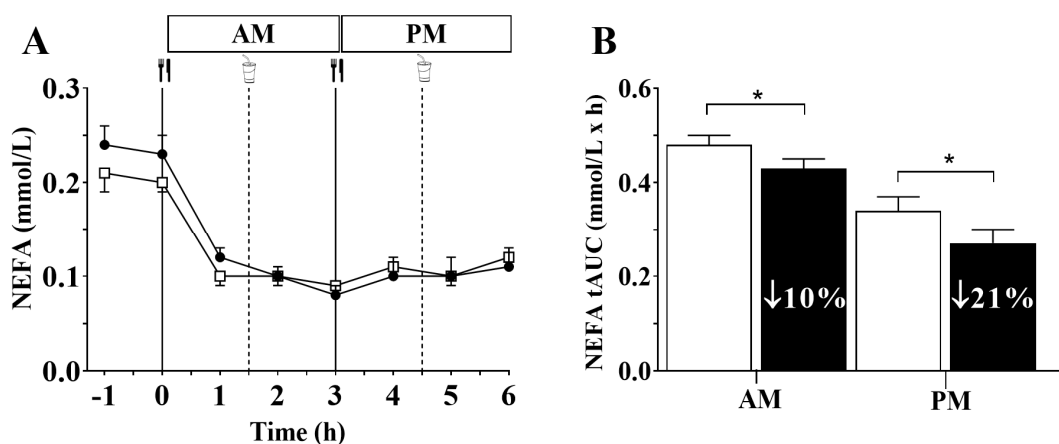


Fig. 4 Fasting and postprandial NEFA concentrations measured during water (open square) and SSB (closed circle) conditions (A). Solid vertical lines indicate timing the breakfast (0 h) and lunch (3.0 h) meals. NEFA total area under the curves (tAUC) responses per trial condition in the morning (AM; 0-3 h) and afternoon (PM; 3-6 h) [Water (white bars) and SSB (black bars) (B)]. Values within the bars indicate the percentage change compared to the water condition. All data are presented as mean \pm SEM. * Difference between water and SSB condition ($P < 0.05$).