# An examination of seagrass monitoring protocols as applied to two New South Wales estuarine settings

### Md Mustafa Kamal Hossain

B.Sc (Hons.) Agriculture (BAU), M.Sc. in Environmental Studies (UNSW)

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**Faculty of Arts and Sciences** 

Australian Catholic University Research Services, Locked Bay 4115 Fitzroy, Victoria 3065 Australia

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### **STATEMENT OF ORIGINALITY**

This thesis contains no material published elsewhere or extracted in whole or in part from a thesis by which I have qualified for or been awarded another degree or diploma. No other person's work has been used without due acknowledgement in the main text of the thesis. This thesis has not been submitted for the award of any degree or diploma in any other tertiary institution. All research procedures reported in the thesis received the approval of the relevant Ethics/Safety Committees (where required).

M.M. Kamal Hossain Date:

## ABSTRACT

Many recent studies have sought to monitor health characteristics of seagrasses, including changes in aerial extent, biomass and fish community structure. While these studies have provided important information on the ecology of seagrass communities on southeast Australia, little attempt has been made to subject these sampling procedures to rigorous experimental testing and review. This study employed commonly-used standard methods for sampling seagrass community characteristics in two sites in New South Wales. Where possible, sampling protocols were tested for accuracy and efficiency at a range of temporal and spatial scales.

The ARCView Geographic Information System was used to construct vegetation polygons of seagrass distribution on the Tweed River, and in the Ukerebagh Channel annually over a 5-year period. For one year (2000), distributions from identical photographs were mapped twice to identify procedural errors. In general, errors relating to incorrect boundary identification were low compared to inter-annual variability. Inter-annual variability in seagrass beds was higher than for adjacent mangrove and saltmarsh.

Estimates of biomass were derived from standard replicate 0.25m x 0.25m quadrats. The experiment contrasted two sites of similar geomorphic setting. Ukerebagh Channel on the Tweed River, and Woolooware Bay with Botany Bay are both shallow, sandy marine deltaic settings supporting stands of *Zostera capricorni*. Significant differences were found in the degree of replication required to identify significant changes in seagrass biomass at the two sites. Ukerebagh Channel supported relatively dense stands of *Z. capricorni* with low intra-site variability. Here 8 replicates were sufficient to detect 10 percent change. Towra Point presented a contrast, in which 15 replicates were required to detect a similar level of change. Woolooware Bay at Towra Point has suffered from increased sedimentation relating to alterations in current velocities at Towra Point, and the result highlights the greater degree of replication required to determine significance changes in disturbed systems.

The fish populations in the seagrass at Towra Point were sampled using buoyant pop nets. Fish communities differed significantly from those sampled in adjacent mangrove and saltmarsh. Differences in fish assemblages between spring high tides, neap high tides and low tides are attributed to movements of fish between seagrass and adjacent mangrove and saltmarsh. This mosaic of habitats is utilized by a number of species over a tidal cycle, with seagrass providing an important low-tide refuge for many species utilizing mangrove and saltmarsh at high tide. Limitations in the efficiency of buoyant pop nets were exposed in a novel experiment which demonstrated differences in escape rates between species. Flat-tailed mullet (*Liza argenta*) are likely to be under-represented in experiments using this technique.

Recommendations are made regarding optimal sampling protocols for monitoring seagrass in the region. All techniques tested are suitable, though some require modification. Some texts have under-estimated the degree of replication required to appropriately monitor changes in seagrass biomass in disturbed systems, where density is lower and intra-site variability higher. The buoyant pop-nets may require modification in open-water seagrass situations where escape by *Liza argenta* and *Acanthopagrus australis* were at unacceptable levels.

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### **Chapter One: introduction and literature review**

#### **1.1 Introduction**

Seagrass communities are a key component of coastal ecosystems through the world (Kikuchi, 1980; Larkum, 1976; West, 1983; Robblee and Zieman, 1984; Walker, 1989; Hutomo and Peristiwady, 1996). Aside from their intrinsic value, they provide food, habitat and shelter for many commercially important species of fish and crustaceans (Poiner *et al.* 1989; Gray *et al.* 1996). For example, in Botany Bay the juvenile fish take shelter in *Zostera capricorni* beds but soon migrate to *Posidonia australis* beds, where the longer leaved meadows and associated epiphytes are considered to provide more food (Middleton *et al.* 1984). Rotherham (1999) found a similar situation occurred in *Z. capricorni* and *P. australis* beds in Port Hacking and St. Georges Basin. Two fish species, luderick (*Girella tricuspidata*) and tarwhine (*Rhabdosargus sarba*), appeared to move to *P. australis* after spending initial stages in *Z. capricorni* meadows.

Seagrasses contribute to detrial food chains by their high rates of primary production (King, 1981). For example, annual primary production of *P. australis* leaf material in Botany Bay was 0.3–2 tonnes/ha/year (West and Larkum, 1979) and in Port Hacking was 2.4 tons/ha/year (Kirkman and Reid, 1979). Seagrasses stabilise sediment, and contribute to the quality of marine and estuarine waters (Gambi *et al.* 1990). The plants filter nutrients and contaminants from the water, act as dampeners to wave action (Short and Coles, 2001).

Seagrasses are aquatic flowering plants and are generally found in soft sediments. In NSW it has been noted that *P. australis* generally occurs on sandy substratum, whereas *Zostera* species grows on a range of substrates (Harris *et al.* 1979; West and Larkum 1983). Seagrasses grow below the high tide level in the sheltered shallow waters of estuaries. They need light to grow and are usually restricted to the upper two meters of water where there is sufficient light (NSW Fisheries, 2004). Minimum light requirements for seagrasses appear to be higher than terrestrial plants (Abal *et al.* 1994). The same

species of seagrass can often be found distributed at different depths in different estuaries, in response to variations in turbidity of the waters. For example, Larkum (1976) observed that the lower limit for *P. australis* at Towra Point, Botany Bay was 3m below mean low water, whereas at Greenpatch, Jervis Bay, the lower limit was 9m. *Zostera* spp. are able to survive at much greater depths in clear waters than they are in highly turbid waters, with depth limits ranging from 2m to 30m (Kennish, 1986).

Seagrasses are composed of a set of vegetative modules: a piece of rhizome, which can be either horizontal or vertical; a bundle of leaves attached to the rhizome; and a root system. The leaves are either strap-like or paired ovals. The leaves of *Zostera* spp. are linear like and 1-5 mm wide and up to 500 mm long. *Posidonia* spp. have strap-like leaves 6-14 mm wide and 300-600 mm long. The leaves of *Halophila* spp. are paired and oval shaped, 5-20 mm wide and 10-50 mm long. The rhizome diameter of *Zostera* species varies from 1.6 - 3.5 mm, *Posidonia* species 5.5 - 9.7 mm and the *Halophila* species of the sexually and asexually, and thus posses two mechanisms by which they can recover from damage (Grey and Moffler, 1978).

In *Zostera* alternate male and female flowers are held in a spathal sheath on up-right reproductive stems produced between spring and late summer. *Zostera* is found growing under a variety of conditions, and exhibits a plasticity in leaf shape to suit. For example, in inter-tidal situations leaves may be as little as 20 mm long and 1 mm wide, whereas when growing in channels with strong currents the leaves attain lengths of more than 500 mm and width of 5 mm. *Zostera* is found in marine tidal waters and brackish non-tidal waters on sandy, muddy and more rarely gravelly sediments. In oceanic waters, on sand or sand-mud, it is often found at the inter-tidal and sub-tidal extremities of the *Posidonia australis* meadows (West, 1983). *Zostera capricorni* is common in New South Wales water, *Zostera muelleri* is thought to occur in infrequently opened lagoons and at some marine locations, south of Sussex inlet.

#### **1.2 Seagrass species in Australia**

Australia has a unique and rich seagrass flora with more than half of the world's species (Hartog, 1970). Seagrasses are a prominent feature of tropical and temperate coastlines of Australia. Australia's 32,000 km coastline contains the largest and most diverse seagrass assemblages in the world (Walker *et al.* 1999). There are 22 species in temperate waters and 15 in tropical waters in Australia. The highest species diversity occurs in Western Australia (CSIRO, 1997).

Globally, there are 58 recognised species of seagrasses (Kuo and McComb, 1989). Most countries have between two to six species. More than half of the world species of seagrasses grow in Australian tropical and temperate waters. Australia is therefore unique in having the largest and most diverse mix of seagrass species in the world (Larkum *et al.* 1989). There are eight species of seagrasses in NSW including *Posidonia australis*, three species of *Zostera* and three species of *Halophila* which are difficult to differentiate (Robertson, 1984). There are a number of species of *Ruppia* in NSW although *Ruppia megacarpa* is considered to be dominant (Robertson, 1984).

Distribution of seagrasses is influenced by a number of environmental factors, many of which are a function of the location within the estuary and the type of estuarine system. Each species of seagrass is distributed according to the interaction of light and temperature, sediment and nutrients, salinity, wave and current action (Day *et al.* 1989). The distribution of seagrasses in Australia is shown in Figure 1.1.



Figure 1.1: Seagrasses in Australia: indicative distribution and sites of disturbance in continental Australia. Source: Environment Australia, 2000 and Kirkman, 1997.

#### 1.3 Seagrasses in Southeast Australia

West *et al.* (1985) mapped the distribution of seagrasses, mangroves and saltmarshes in NSW estuaries from aerial photos taken between 1977 to 1984, and reported 155 km<sup>2</sup> of seagrass in 111 estuaries. The largest single area of seagrass in NSW was found in Wallis lake, which at that time contained 21% of the state's total seagrass. While no more recent estimates on a state-wide basis exist, there have since been some significant changes (decrease or increase) in the estuaries of NSW (Poiner & Peterken, 1995; NSW Fisheries, 1999; Meehan, 1997 and Williams and Meehan 2001). The distribution of seagrasses in southeast Australian seagrasses is given in Table 1.1.

Table 1.1. Distribution of seagrass species in southeastern Australia (Tweed River to Corner Inlet) after West *et al.* (1989).

Genus	Species	Distribution in S. E. Australia		
Zostera	capricorni Aschers	Whole coast		
Zostera	mulleri Irmisch & Aschers	Jervis Bay and southward		
Heterozostera	tasmanica	Port Stephens and southwards		
Halophila	decipiens Ostenfeld	Whole coast		
Halophila	ovalis (R.Br.) Hook.f.	Whole coast		
Halophila	australis Doty & Stone	Central coast and southwards		
Posidonia	australis Hook.f.	Wallis Lake and southwards		
Ruppia	megacarpa	Whole coast		

Source: Watford and Williams, 1998

Along the New South Wales coast seagrasses are confined to estuaries such as Botany Bay, except for the large sheltered embayment of Jervis Bay (CSIRO, 1997). The majority of the coastal features where seagrass grows in New South Wales are estuarine. *Zostera* spp. occur in 93% of estuaries, *Halophila* spp. in 66% and *P. australias* in 43% (West, 1983). There are also several open bays in New South Wales containing seagrasses. Bays which experience high wave energy are unlikely to support *P. australis*, eg. Batemans Bay (West *et al.* 1989).

Botany Bay is a shallow, marine dominated open embayment, located 12 km south of Sydney, NSW. It was reported (West *et al.* 1985) as having 340 ha of seagrass, the eighth largest seagrass area in New South Wales. Principal seagrass species are *P. australis* and *Z. capricorni* with smaller beds of *H. ovalis* and *H. decipiens* (Larkum, 1976).

#### **1.4 Threat / losses of seagrass**

Seagrasses can be easily destroyed. Once seagrass meadows are damaged, their recolonisation can be very slow. The leaves of the seagrass grow quickly, the rhizome grows relatively slowly. The ability to recover after disturbance varies between seagrass species. In NSW, observations over many years have indicated that *Halophila* spp. are relatively quick to respond to change and may recover from damage within months, while *Zostera* spp. take several years to recolonise denuded areas (West and Larkum, 1983; Larkum and West, 1990 and West, 1990). Larkum (1976) cleared seagrass from small plots ( $4m^2$ ) in Botany Bay and assessed regrowth, three years no regrowth was observed and movement of rhizomes into the denuded areas was negligible. Meehan (2002) estimated the rate of recolonisation of *P. australis* within blast holes in Jervis Bay, the lateral rate of recovery was estimated at 250 mm per year, and complete recovery may take nearly a century.

There has been a noticeable decline in the extent of some inshore habitats, particularly seagrasses in NSW. It was that at least 50% of NSW's seagrasses has been lost between the 1980s and 1990s (Poiner and Peterken, 1995). The condition of seagrasses in NSW is highly variable, but in general seagrass beds near urban or industrial area are often degraded, while more isolated seagrass areas are in good health (NSW Fisheries, 1999b). Overall, NSW has lost an estimated 50% of the *Zostera* sp. seagrass in its estuaries since European settlement (Zann, 1995). This loss has occurred through both natural events (eg. cyclones, floods) and anthropogenic events (eg. nutrient enrichment, coastal development etc).

Activities that threaten seagrass habitat include change to drainage, land management practices which lead to sedimentation, direct removal of seagrasses by dredging, boating, fishing or sand mining, nutrient run off from agricultural land and aquaculture, point source discharge from industry and storm water drains, introduced pests and diseases (Butler and Jernakoff, 1999). Contamination by herbicides, petrochemicals and heavy metals from catchment run-off has been linked with massive die-back of seagrasses in many areas. Changes to hydrology for harbour and coast line development have also

contributed to a decline in seagrass beds around the major population centres (SoEAC 1996). Sand deposition and cropping were thought to be the cause of the loss of *Z*. *capricorni* meadows in Moreton Bay, Australia (Kirkman, 1978).

Seagrasses are also susceptible to the effects of natural climatic events such as cyclones, flood and storms. Preen et al. (1995) reported that flood and cyclone related loss of more than 1000 km<sup>2</sup> of seagrass from Hervey Bay, Queensland. They found that seagrass loss in shallow water (<10m) could be attributed to uprooting during heavy seas; however seagrass loss in deeper water was probably due to a persistent plume of turbid water resulting from flooding or developed catchments. In Botany Bay mapping from aerial photographs taken between 1942 and 1986 showed that more than half (about 57%) of the seagrass was lost (Larkum and West 1990). Seagrasses in the Gulf of Carpentaria sustained considerable damage due to the passage of Cyclone Sandy in 1985. The cyclone removed, undermined or smothered 70% (128 km<sup>2</sup>) of seagrasses in the area and scouring and smothering eventually destroyed the remainder of the bed. Overall this represented a 20% loss of the seagrasses in the Gulf (Poiner et al. 1989). In another example, Seddon et al. (2000) reported a loss of 12,717 ha of intertidal and shallow subtidal Amphibolis antarctica and Zostera spp. in northern Spencer Gulf, South Australia. Increased turbidity brought by foreshore and catchment development appeard to reduce the depth limit of P. australis in Botany Bay (Larkum, 1976; Larkum and West 1990). Construction of airport runways and port facilities also caused the decline of seagrasses in this area.

#### **1.5 Monitoring of seagrass**

Seagrasses play an important role in estuarine ecosystems by stabilizing sediments and providing food and shelter to a wide variety of aquatic organisms. It is therefore important to monitor the changes of distribution and health of seagrasses so appropriate management strategies can be adopted. Monitoring is also important to ensure that any deterioration in seagrass health is quickly detected and to prevent large scale losses occurring. Monitoring can be carried out using remote sensing and on *in situ* techniques. Satellite imagery has until recently been provided at a scale which precluded the

detection of small scale changes in seagrass health. Aerial photographs cannot readily detect changes in density or biomass of seagrass. Aerial photograph also may be compromised by sun light, shading, cloud cover or wave action. Further, the process of on-screen digitizing can introduce a number of inconsistencies and errors which might contribute to variation in measures of seagrass area and distribution.

Monitoring the health of seagrass *in situ* usually requires describing changes in a specific seagrass community using one or more variables. The selection of a variable to monitor can be subjective (Wood and Lavery, 2000). The variables that have widely used to detect small scale changes include biomass, density and productivity (Kirkman, 1996). There is no standard set of protocols to monitor seagrass health using any of these variables, although there have been some attempts to evaluate various monitoring techniques (Mellors, 1991; Inglis and Smith, 1995). Many of the studies that have monitored seagrass health in southeast Australian estuaries have varied in terms of the type of variables measured, the number of samples, replication and the sampling interval (Larkum, 1976; West and Larkum, 1979; West, 1990; Wolterding, 2000).

#### 1.6 Examination of fish utilizing seagrass habitat

Seagrass meadows have characteristics that make them a suitable habitat for many species of animals. The three-dimensional structure of the vegetation, with its network of roots and rhizomes and often dense leaf canopy, offers hiding places that protect against predation and also provides substrate for attachment (Hemminga and Duarte, 2000). The fish community of seagrass meadows is not fixed or constant for any particular species or combination of species. Juvenile species often dominate the fish communities of seagrass beds. In a study of the fish communities of estuarine seagrass beds in northern Queensland, 134 species were recorded (Coles *et al.* 1993). The average length of the individual species caught was only 32 mm because most of the fishes were juveniles, although some large adult fishes with length over 60 cm were also present (Coles *et al.* 1993). In the case of the seagrass meadows in Cairns Harbour, Queensland it was calculated that the potential annual yield of the three major commercial prawn species

from these meadows amounted to between 0.6 and 2.2 million Australian dollars per year (Watson *et al.* 1993).

Connolly (1994) compared fish catches from pop and seine nets in shallow eelgrass meadows to determine the relative catching efficiencies of the two methods. Buoyant pop nets usually consist of four mesh walls, quickly released and can be lifted clear of the water to collect ensnared fish. A total of 15 species were collected by Connolly (1994) and the number of fish caught in pop nets was greater than that of seine nets.

Measures of seagrass health or condition should consider not only the structural attributes of the seagrass (density, biomass, distribution) but also the habitat values of the seagrass. Bouyant pop-nets have been used by Connolly (1994) and Mazumder *et al.* (2004) to determine fish assemblages in shallow vegetated estuarine habitats. While these nets have provided insights into fish assemblages in these environments, the catch efficiency of the technique remains largely untested.

#### 1.7 Aims and structure of the thesis

The thesis seeks to contribute to the development of accurate methods for determining and monitoring seagrass condition by testing the most commonly used methods of seagrass condition assessment.

Chapter 2 uses the ARCView Geographic Information System to investigate the utility of on-screen digitizing of georectified air photographs as a means or accurately determining seagrass distributions. For this exercise, the Tweed estuary was used because of the clarity of the water and the extensive air photographic record. Maps were produced for the Ukerebagh Channel, one of the most important seagrass habitats on the Tweed. A variety of scales of temporal replication were considered to determine optimal mapping periods for change detection. Errors associated with on-screen digitizing were assessed by repeat mapping for the year 2000.

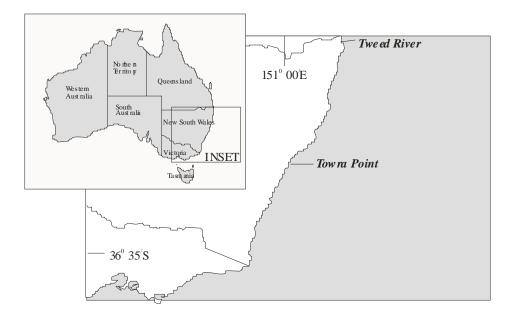


Figure 1.2: Location of Towra Point (Woolooware Bay) and Tweed River within New South Wales

Seagrass biomass is commonly estimated on the assumption that a design a 0.0625m<sup>2</sup> quadrats with 8 replicates in a 15m X 20m plot will provide an accurate indication of biomass and that such a measure represents a useful as a benchmark against which change can be detected. This assumption was tested by applying this protocol seasonally over a period of 12 months in two estuarine setting. The two settings were geomorphologically similar. Both the Ukerebagh Channel on the Tweed River and the Woolooware Bay of Botany Bay being marine sand in a shallow estuarine deltaic settings (Fig. 1.2). However, the two settings differed in the degree of sediment stability and seagrass biomass, with the Woolooware Bay seagrass receiving sediment influx which created a patchy distribution. This afforded the opportunity to assess the sampling protocols in stable and disturbed settings. A power analysis was used to determine the degree of replication which would be required to detect appropriate levels of change in each context. The power analysis formed the content of Chapter 3.

Finally, it is important to asses the efficiency of methods used to sample fish assemblages in seagrass, because seagrass condition can also be assessed on the basis of functionality (e.g. fish habitat) rather than structure alone. Fish were sampled at Woolooware Bay using pop nets, monthly for a period of 12 months. The sampling strategy took into consideration linkages between seagrass and adjacent mangrove and saltmarsh habitat, by sampling at high and low tide, and in conjunction with a separate project assessing fish utilization of the adjacent mangrove and saltmarsh. In keeping with the theme of the thesis, an effort was made to determine the efficiency of the buoyant pop net by releasing fish into the sprung net and measuring recapture rates for a range of species. This forms the content of Chapter 4.

The concluding chapter (Chapter 5) provides recommendations about optimal strategies for measuring seagrass community characteristics, both floral and faunal, and detecting change. The aim of the work is therefore to provide guidelines for the more appropriate and reliable monitoring of this important coastal habitat.

### **Chapter Two: seagrass mapping**

#### 2.1 Introduction

Remote sensing is a technique that measures the reflectance characteristics (for example, colour, electromagnetic radiation) of the earth's surface and relates these measurements to vegetation and land use. Remote sensing of seagrass communities using aerial photographs has been used to document the status (West *et al.* 1985) and changes (Larkum and West 1990, Meehan 1997) in seagrass cover in a number of estuaries in southern New South Wales. Aerial photographs have been used to document the status and examine long term changes in seagrass cover in a number of estuaries in southern NSW (Larkum and West, 1990; Williams and Meehan 2001, Meehan 2002). These studies have been able to determine whether seagrass communities have decreased or increased over a period of time.

Seagrass meadows form characteristic landscapes that are visible from remotely sensed imagery (Steffenson and McGregor, 1976; Kendrick *et al.*, 1999 and 2000; Mumby and Green 2000b). Different species of seagrass, such as *Z. capricorni* and *P. australia* can exhibit noticeable differences in colour, tone and texture that can be detected on present and past remotely sensed imagery (Larkum and West, 1990; Meehan, 1997; Watford and Williams 1999).

#### **Case study**

The Tweed River Entrance Sand by Passing Project is a joint project under taken by the State Governments of Queensland and New South Wales in conjunction with the Gold Coast City Council and Tweed Shire Council. The main aims are to establish and maintain a navigable entrance to the Tweed River and to enhance and maintain the amenity of the Southern Gold Coast beaches.

The Tweed River is located in NSW near the NSW/Queensland border just south of the Gold Coast. The project uses a pier mounted jet pump system to bypass sand around a trained river entrance. The Tweed River entrance is in an area with high wave energy and longshore sediment transport averaging 500,000 cubic metres per annum (Hyder *et al.* 1997). During the period 1 to 25 March 2001, modeled daily sand transport rates varied from 442m<sup>3</sup>/day to 24,696m<sup>3</sup>/day (McQuade and Jackson, 2001). There have been two stages in the project : from 1995 to 2001 a temporary sand by-pass system and from 2001 a permanent system was installed. During the temporary phase 3.6 million cubic metres of sand were dredged from the entrance and used to nourish the beaches to the north (Dyson *et al.* 2001).

The aims of this study is

- to determine whether seagrass extent in Ukerebagh Channel had undergone change following the opening of the Tweed sand bypassing project
- to conduct annual mapping of seagrass community distribution in the Ukerebagh Channel to determine inter-annual variability in seagrass extent and the optimal frequency of mapping for change detection
- 3) to determine the extent of digitizer error.

#### 2.2 Study site selection

As an exercise in developing monitoring skills a small portion of a NSW estuary was examined for temporal change in seagrass distribution. The Tweed River was chosen for the study as the sand bypassing project, provided an opportunity to examine seagrass variability in relation to a pulse event. Good air photo coverage exists prior to and following this event. Clarity of water is excellent for all photograph year.

#### 2.3 Method

Aerial photographs are the most commonly used form of remotely sensed data for high resolution seagrass mapping studies. Aerial photographs of Tweed River, and successive years for the Ukerebagh Channel, were obtained from the Department of Infrastructure Planning and Natural Resource (DIPNR). All obtained images are not in same quality.

Image quality depends on image resolution, air quality, weather conditions, clarity of the water and tide levels.

The photographs were visually interpreted and the seagrass beds identified on the basis of reflectance characteristics (colour, tone and texture). The observable characteristics of seagrass species are given in Table 2.1.

Community type	Observable characteristics				
Seagrass Spp.	Colour range light to dark green or light to medium				
	Black. Colour varies depending on depth. Beds are				
	large, smaller and fragmented. Sparse beds may be				
	lighter in appearance. Beds usually have soft edges				
	and are smooth in appearance.				

Table 2.1: Observable characteristics of seagrass species in estuaries of NSW

Ground truthing, previous maps and knowledge of seagrass ecology were also used to identify the seagrass beds and draw their boundaries. The full list of aerial photographs used is presented in Table 2.2.

Aerial photographs were scanned at 360 dpi (dots per inch) and saved in JPEG file format and imported into the ArcView Geographic Information System (ESRI Inc). They were georectified to correct distortions of scale caused by the varying distances of photographed objects from the camera lens. A minimum of 6 ground control points were used to rectify each image, all derived from recognizable fixed points on the CMA topographic maps of the region.

The on-screen digitizing functions of ArcView were used to create polygons of seagrass areas. The size and shape of the polygons within or between sets of photos vary according to a number of factors, including the scale of the photo and the complexity of the habitat (Green *et al.* 2000b). For example a fragmented seagrass habitat may need a large number of smaller polygons than a continuous seagrass bed. The entire seagrass

was mapped on two occasions, before and after the beginning of the Sand Bypassing project. The following criteria were used when mapping seagrass communities :

-Seagrass beds were differentiated on the basis of colour, tone, texture and geomorphic and geographical context, using the criteria for saltmarsh and mangrove set out by Saintilan and Wilton (2000).

-Interpretations were crossed-checked with the earlier surveys of West et al. (1985).

-Where aerial photographs were compromised by solar reflectance, insufficient light penetration or shading by other vegetation, mapping was not attempted.

Table 2.2: Aer	ial photographs	used to	map	the	distribution	of	seagrass	species	in
Ukerebagh Chan	nel, Tweed River	r.							

Date	Photo series	Run	Print number
28/05/01	Beach Protection Authority Tweed heads to Currumbin	Run 3	100, 101
11/05/00	Beach Protection Authority Tweed heads - Currumbin	Run 3	178
19/10/99	Beach Protection Authority Tweed heads to Currumbin	Run 3S	104,105
7/5/98	Beach Protection Authority Tweed heads – Currumbin	Run 3 Run 1	105, 112
10/09/97	Beach Protection Authority Tweed heads - Currimbin	Run 3	137, 146

#### Ground-truthing of intertidal seagrass communities

Ground-truthing is the process of determining the accuracy of mapped vegetation units by comparison with field observation. Field surveys were carried out to identify the status (sparse or dense) and boundaries of seagrass beds and to assist in visual interpretation of the aerial photographs. Ground-truthing was conducted in 28 randomly selected locations in the Ukerebagh Channel. A bathscope was also used to look at the seagrass and its boundaries in deeper water. This is particularly important if different species of seagrass were present with similar textures and colours in the air photograph, and the geomorphic context did not allow easy differentiation.

#### **Mapping Error**

Some component of the estimated changes in the extent of seagrass will be due to variations in quality between the fine set of photos (Table 2.2), georectification error and digitizing error. Variations always occur in photograph quality and image clarity leading to errors in aerial photography analysis and interpretation. Digitising error is also a potentially significant source of error. To determine the contribution of digitizing error, seagrass boundaries were mapped twice for the photo (only one: Run 3, Print 178) in 2000.

#### 2.4 Results

Seagrass extent increased in the period immediately following the introduction of the Tweed sand bypassing project. The area of seagrass increased from 61.2 hectares in 2000 to 66.77 hectares in 2000 (Figures 2.1 and 2.2).

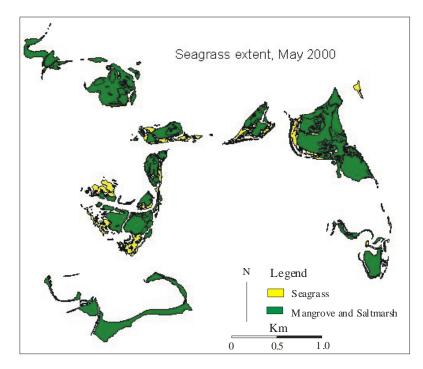


Figure: 2.1 Distribution of seagrass and other intertidal wetlands in the lower Tweed River in 2000.

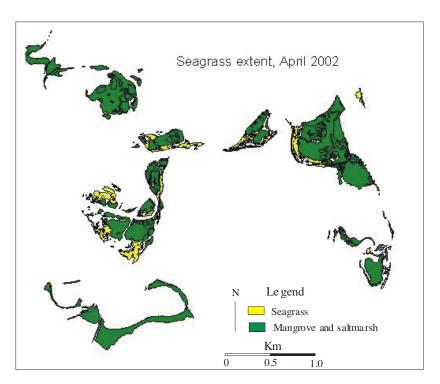


Figure: 2.2 Distribution of seagrass and other intertidal wetlands in the lower Tweed River in 2002.

Changes in seagrass extent within Ukerebagh Channel between 1997 and 2001 are shown in Figures 2.3 to 2.8. The seagrass cover reached its highest point in 2001 (8.95 ha). The aerial cover is in tables 2.3 and 2.4.

Year	Areas (ha)	
1997	7.04	
1998	7.47	
1999	7.05	
2000(a)	7.90	
2000(b)	8.12	
2000 (mean)	8.01	
2001	8.95	

Table 2.3 : Area of Seagrass vegetation in Ukerebagh Channel, Tweed River

Table 2.4: Seagrass cover in percentage (increase or decrease) in Ukerebagh Channel

Year	Percent (%) change	
1997-1998	6.10 increase	
1998-1999	5.62 decrease	
1999-2000	12.05 increase	
2000a-2000b	2.9 increase	
2000-2001	13.29 increase	

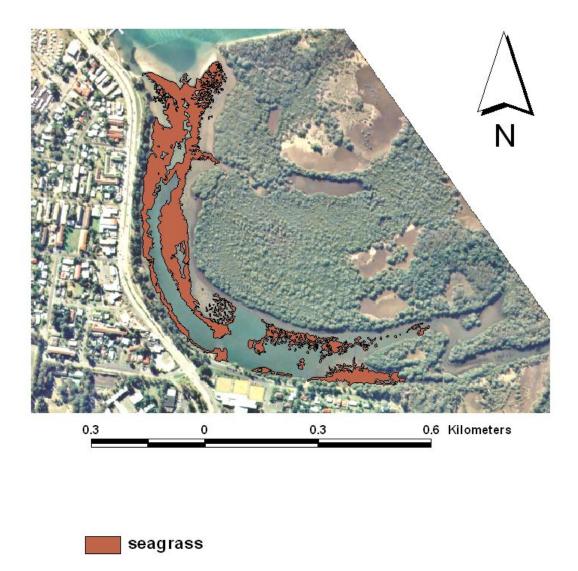


Figure 2.3: Seagrass Extent in Ukerebagh Channel 1997

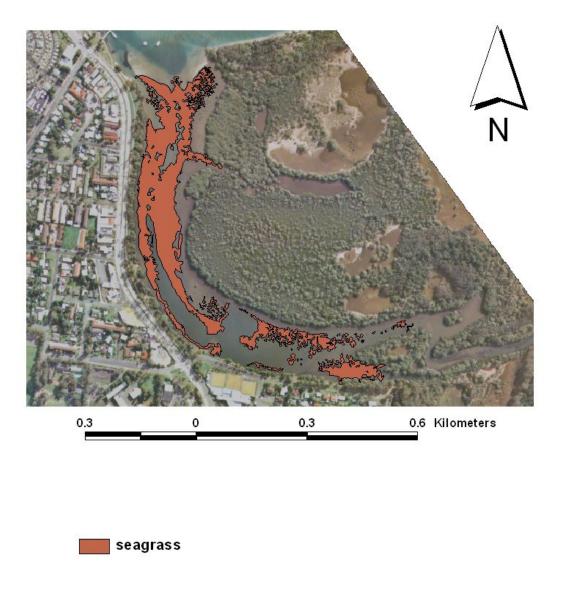


Figure 2.4: Seagrass Extent in Ukerebagh Channel 1998

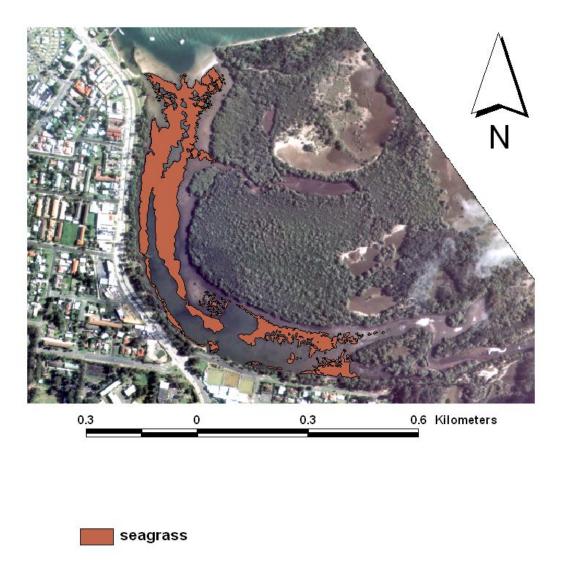


Figure 2.5: Seagrass Extent in Ukerebagh Channel 1999

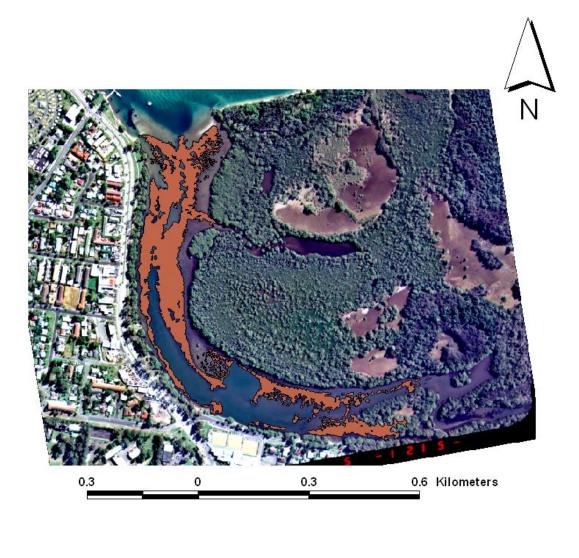




Figure 2.6: Seagrass Extent in Ukerebagh Channel 2000 (a)

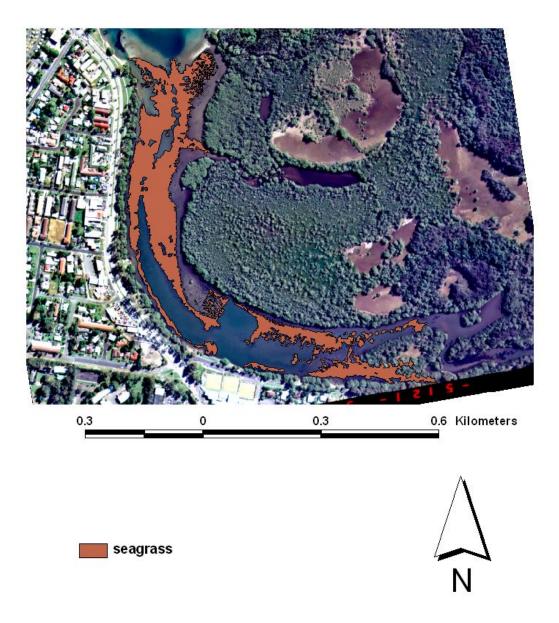


Figure 2.7: Seagrass Extent in Ukerebagh Channel 2000 (b)

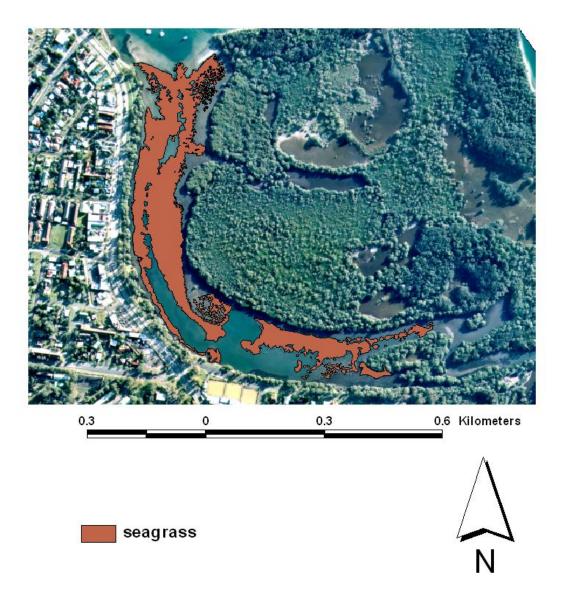


Figure 2.8: Seagrass Extent in Ukerebagh Channel 2001

### **2.5 Discussion**

Seagrass in the Ukerebagh Channel showed a consistent increase in the period of 1999 to 2001. The overall rate of seagrass increase is 6.7% per annum. Factors which promote the increase of seagrass area include water clarity and salinity (which may be associated with drought conditions) and geomorphic stability. There is no indication that the small change in monitored tidal hydrological conditions that may be associated with the Tweed River Entrance Sand Bypass Project within the Tweed River estuary are having a detrimental impact on seagrass beds.

The Tweed River Entrance Sand Bypassing Project, Environmental Impact Statement/Impact Assessment Study (EIS/IAS) predicts very small tidal changes directly attributed to entrance improvements as a result of the operation of the Tweed River Entrance Sand Bypass Project (Hyder 1997). The EIS/IAS predicts that the operation of the Sand bypassing system will reduce the net infeed of marine sand into the river estuary. Operation of the system has the beneficial impact of reducing growth of the lower estuary shoals and lessening their adverse impact on tidal flushing and water quality. To date monitoring of the tidal data shows that there has been no significant change in tidal conditions that can be contributed to the operation of the sand bypassing system.

Meehan (2002) stated that in southern NSW, the area of *P. australia* has declined in five out of six estuaries over the last fifty years. In Port Hacking *P. australis* communities declined 18% in 1951-1999, in St.George Basin seagrass cover declined by 27% from 1961-1998. In Wagonga Inlet seagrass cover declined slightly (10%) from 1957- 1994. In Bermagui River seagrass cover declined by 33% from 1957 –1998. These declines occur due to a number of anthropogenic impacts, including engineering works, dredging and deterioration in water quality. Natural impact, such as storm damage, may have occurred at some sites.

By contrast, the Tweed River *Zostera* community distributions are fairly stable, at least over the time period of this study. There were no significant change in seagrass extent following the Tweed Sand Bypassing Project – a slight increase was noted. Within the Ukerebagh Channel, seagrass extent appears to have increased over a five-year period, in the order of 5-10 % per year. Given that the degree of error calculated by the remapping of 2000 was relatively small (2.9%), most of the increase is assumed to be real increases in seagrass cover. This would suggest that annual measures of seagrass extent can generate useful indications of change over that timescale, in contrast to mangrove and saltmarsh where five-year surveys are recommended (Saintilan and Wilton 2000).

# **Chapter Three: sampling biomass of seagrass**

## **3.1 Introduction**

Seagrasses are an important and often dominant component of ecosystems in estuaries and shallow coastal waters of Australia (Larkum, 1977, Long et al., 1994). Their roles are so important that seagrass meadows are considered to be the most valuable ecosystems in terms of the value-added benefits of the services they provide (Costanza et al. 1997). Studies in other parts of the world have shown that many seagrasses have high primary production and that they may support rich communities, including many commercially important animals (Thayer et al., 1975, Smith, 1981; Charpy-Roubaud and Sournia, 1990). They provide nursery grounds, adult habitat and food for commercially important prawns and fish (Young, 1978, Bell and Pollard, 1989, Blaber et al., 1992), wading birds and dugongs (Preen et al., 1992). They also produce large amounts of detritus and dissolved organic matter (Moriarty et al., 1984). Seagrasses are areas of high food abundance (Hill and Wassenberg, 1993). They are present in only 0.15% of the ocean's surface (Charpy-Roubaud and Sournia, 1990) and contribute 1% of the net primary production of the global ocean (Duarte and Cebrian, 1996). In Botany Bay seagrasses are a major source of detritus (West and Larkum, 1979). Reliable and accurate estimates of the seagrass biomass are essential for estimating production and the links with the ecological system components that rely directly or indirectly on this resource.

Within Australia, only a few studies have considered in detail the biomass of *Z. capricorni*. In NSW biomass data were available for the Tuggerah Lakes (100 km north of Sydney) (Higginson, 1965), and in Queensland, Moreton Bay, (Kirkman, 1978). Harris *et al.* (1979) studied seasonal changes in biomass and phenology of communities in Illawarra Lake (110 km south of Sydney) and Kirkman *et al.* (1982) studied the biomass, leaf growth and productivity of *Zostera* communities in Port Hacking (30 km south of Sydney). Larkum *et al.* (1984) studied the standing stock, growth and shoot production of *Z. capricorni* in Botany Bay, New South Wales and Kerr *et al.* (1990) studied the

seasonal changes in standing crop of Z. muelleri in south-eastern Australia. No studies have been conducted of biomass and standing crop of Zostera sp. in the Tweed River, in spite of the potential of significant engineering works at the mouth of the Tweed to disrupt seagrass beds.

## Aims of this study

 $\Delta$  The current experiment investigated the seasonal variation in the abundance of the above ground biomass of *Zostera* sp. in two locations within NSW : Woolooware Bay segment of Botany Bay and Ukerebagh Channel in the Tweed River.

 $\Delta$  Relationships between season, seagrass biomass and optimum number of quadrats for sampling were also examined.

 $\Delta$  The primary aim of this investigation was to determine the efficacy of commonlyproposed sampling regimes for seagrass biomass.

A number of authors (Larkum *et al.* 1984; Larkum and West, 1990; Kirkman and Cook 1982; Mellors 1991, and Lanyon and Marsh (1995)) suggest that the most appropriate level of sampling is 0.25m x 0.25m quadrats. The level of replication has commonly between 10 replicates plot (eg. Larkum *et al.* 1984). The opportunity exists to apply Power Analysis to assess the sufficiency of this level of replication in two estuarine settings.

Table 3.1: Size of the quadrat used by a number of authors for collecting seagrass sample in different parts of Australia.

Author	year	Location	Species	Quadrat Size (m <sup>2</sup> )
Kirkman and Cook	1982	Port Hacking, NSW	Z. capricorni	0.0625
Larkum etal.	1984	Botany Bay, NSW	Z. capicorni	0.0625
Larkum and West	1990	Quibary Bay and Silver beach, NSW	Posidonia sp.	0.25

Mellors, J.E.	1991	Green Island, North Queensland	H. uninervis 0.25
Neverauskas, V. P.	1987	Gulf St. Vincent, South Australia	Posidonia sp. 0.0625 Amphibolis sp.
Lanyon and Marsh	1995	North Queensland	Seagrass sp. 0.0625

#### 3.2 Study sites

The biomass sampling was undertaken at Woolooware Bay (Towra Point) in Botany Bay (Figure 1.2) and the Ukerebagh Channel in Tweed River estuary (Figure 1.2). Botany Bay (34°00′ S, 151°11′ E) is a marine dominated open embayment, 12 km south of Sydney, New South Wales. Among the 130 estuaries of New South Wales, the bay was reported in a survey to contain 340 ha of seagrass, the eighth largest seagrass area in New South Wales (West *et al.* 1985). Tweed Heads is located 860 km north of Sydney. Both sites have some geomorphological similarities, being sandy marine deltas. The principal seagrass species in Botany Bay are *Posidonia australis* and *Zostera capricorni*. (Larkum and West, 1990). Sample sites Woolooware Bay segment of Botany Bay and Ukerabagh Channel in Tweed Heads were a monospecific stand of *Zostera* sp. Both sites are important as a commercial and recreational fishing area, as a site of historic significance and a popular recreation area in terms of boating, fishing, bush walking and bird watching.

#### **3.3 Sampling Methods**

The sampling method for estimating the biomass utilises random samples within plots. To improve precision, Downing and Anderson (1985) recommended the use of small quadrats with many replications in preference to a few large quadrats. For example Larkum *et al.* (1984) used ten (0.25m X 0.25m) 0.0625m<sup>2</sup> quadrats, for collecting *Zostera capricorni* sample from Botany Bay. Larkum and West (1990) collected *Posidonia* samples for biomass estimation from Quibray Bay and Silver Beach, Botany Bay, using six 0.25m<sup>2</sup> quadrats. Kirkman and Cook (1982) collected *Zostera capricorni* from Port

Hacking for estimating biomass through ten  $(0.25 \text{ m X } 0.25 \text{ m}) 0.0625 \text{ m}^2$  quadrats. Mellors (1991) used  $0.25 \text{ m}^2$  quadrats for collecting sample. Neverauskas (1987) used nine  $(0.25 \text{ m X } 0.25 \text{ m}) 0.0625 \text{ m}^2$  quadrats to collect seagrass sample from the study sites. Lanyon and Marsh (1995) also used sixteen (0.25 m X 0.25 m)  $0.0625 \text{ m}^2$  quadrats for collecting sample.

In each of Woolooware Bay and Ukerebagh Channel 32 of  $(0.25 \text{ m X } 0.25 \text{ m}) 0.0625 \text{ m}^2$  quadrats (8 in each of four plots) were placed randomly within a plot of dimensions 15m X 20m (Figure 3.1). Seagrasses communities were sampled four occasions between winter/2002 until autumn/2003 (four occasions in one year). Sampling was carried out at low tide (< 0.4m) to avoid problems of water turbidity. All plant parts that originated from outside the quadrats were placed outside the frame and those that originated from inside are placed inside the frame. The quadrat was then pushed into the substrate and pegged down to prevent movement while the contents were removed. Above ground plant parts were cut at their bases. Care was taken to ensure that samples were always collected from a previously untouched area of seagrass.

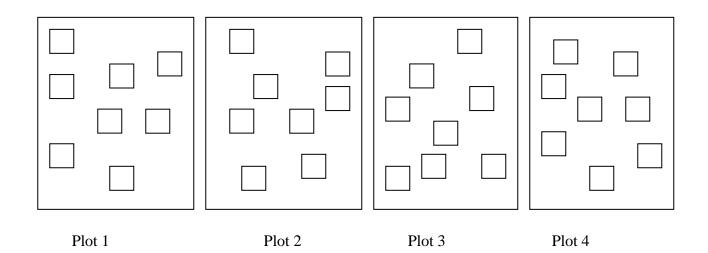


Figure 3.1 Schematic representation of biomass sampling strategy at Woolooware Bay and Ukerebagh Channel, showing plots and replicates within the plots.

## Sampling problems in seagrass beds

Seagrasses often cover large expanses of mudflats which remain partially underwater even at low tide, making sampling difficult. The soft glutinous part often create major problems relating to access and disturbance. The mud is difficult to walk on, algae are abundant and clouds of silt arise to obscure the view during sampling. Some algae are epipelic (on the mud itself), loose-lying or epiphytic (on seagrass leaves), or are on various non-living substrata protruding above the mud.

## **Sample Processing**

Leaves and stems were brought back to the laboratory and rinsed with fresh water to remove the salts and sediments, and the plant parts were then separated from the associated epiphytes. The cleaned leaves and stems were oven dried to constant weight at 80<sup>o</sup>C (Larkum and West, 1990) for 24 hours and weighed immediately since they reabsorb moisture quickly.

#### **Statistical methods**

Univariate analysis (ANOVA) and post-hoc Student-Newmans-Kules (SNK) test were performed using GMAV5 (Underwood and Chapman 1989) to determine the significance of differences of above ground biomass among the seasons in two sites. Two way analyses of variance were used (Underwood, 1997) where season was considered as the first factor with three levels (1=Winter, 2= Spring, 3= Summer, 4= Autumn) that were orthogonal and fixed. Site was considered as second factor with two levels (1=Towra Point, 2=Tweed Head) that were also orthogonal and fixed.

## **3.4 Results**

ANOVA results showed that the dry weight biomass of seagrass at Woolooware Bay in (Towra Point) vary within the season. The above ground biomass among the seasons differed significantly (P<0.0001, Table 3.2).

Table 3.2: ANOVA results for above ground biomass among seasons at Woolooware Bay, NSW. Cochran's Test C= 0.3156

Source of variation	DF	MS	F	Р
Season	3	9.8597	23.13	0.0001
Residual	124	0.4262		
Total	127			

The *post-hoc* SNK test for biomass in different seasons showed that the above ground seagrass biomass in winter is significantly different with the spring. Biomass is also significantly different between winter and summer & autumn and spring. There is no significant difference in biomass between the seasons of summer and autumn. The dry weight in spring is higher than all other seasons (Figure 3.2).

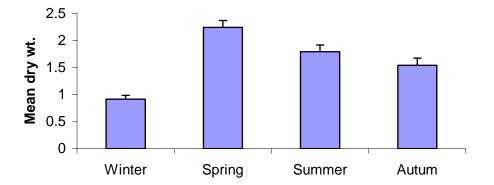


Figure 3.2: Mean dry weight (+ SE ) of seagrass in Woolooware Bay at Towra point, in different seasons 2002-2003: n=128.

In the case of Tweed Head the ANOVA result showed that the biomass also varied within the seasons. Biomass among the seasons differed significantly (P < 0.0003 Table 3.3). Mean dry weight of above ground biomass at Ukerebagh Channel was higher than at Woolooware bay.

Table 3.3: ANOVA results for above ground biomass between the seasons in Tweed Head, NSW : Cochran's Test C=0.4955

Source of variation	DF	MS	F	Р
Season	3	39.1152	6.62	0.0003
Residual	124	5.9110		
Total	127			

The post-hoc SNK test for biomass in different season showed that summer is significantly different with other seasons. Biomass between autumn and winter did not vary significantly. The mean dry weight of above ground biomass in summer is higher than in all other seasons (Figure 3.3).

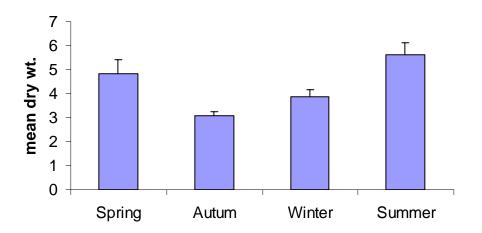


Figure 3.3: Mean dry wt. (+SE) of seagrass in Tweed Head in different seasons 2002-2003: n=128.

Results of the 2-way ANOVA demonstrate significant differences between the biomass of seagrass at Ukerebagh Channel, Tweed River and Woolooware Bay, Towra Point (P<0.001). The significant interaction term (Season \* River: P = 0.004) suggests that the pattern of seasonal variability differs between the two sites (Table 3.4). Towra Point has a more pronounced Spring peak (Figure. 3.2).

Table 3.4: Two-way ANOVA results for factors River (Tweed and Woolooware Bay) and Season (Winter, Spring Summer and Autumn).

Dependent varia					
	Type III				
Source of	Sum of		Mean		
variation	Squares	df	Square	F	Sig.
Corrected	(21.720(a))	7	88.818	28.031	0.000
Model	621.729(a)	1	00.010	28.051	0.000
Intercept	2284.960	1	2284.960	721.129	0.000
Season	103.558	3	34.519	10.894	0.000
River	474.804	1	474.804	149.847	0.000
Season * River	43.367	3	14.456	4.562	0.004
Error	785.809	248	3.169		
Total	3692.497	256			
Corrected Total	1407.538	255			

## **Tests of Between-Subjects Effects**

a R Squared = 0.442 (Adjusted R Squared = 0.426)

## **Statistical Power**

Dependent Variable: Biomass

Power analysis performed on data of mean biomass for seagrass at Woolooware Bay indicated that eight replicate quadrats per plot will only detect significant change of 70% or greater (Figure 3.4). Possibly as many as 15 quadrats per plot would be required to detect a 20% change. Alternatively, power analysis performed on seagrass mean biomass data in Ukerebagh Channel (Tweed Heads) indicated that with eight replicate quadrats is optimal, less than eight quadrats will not detect (as significant) change of less than 10% (Figure 3.5). More than eight quadrats is unnecessary, as statistical power is not increased.

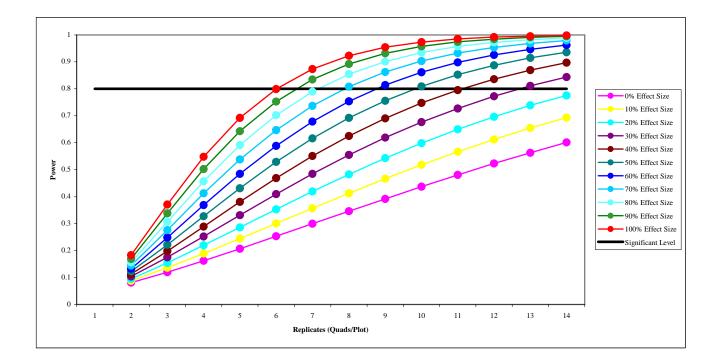


Figure 3.4. Power analysis showing the change detectable with increasing plot replication at Woolooware Bay (Towra Point).

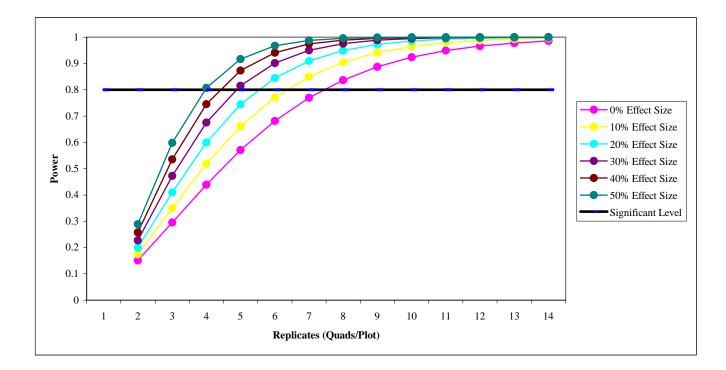


Figure 3.5. Power analysis showing the change detectable with increasing plot replication at Ukerebagh Channel (Tweed Heads).

## **3.5 Discussion**

The current study has demonstrated that seagrass biomass varies significantly between the sites and periods of time (seasons). The differences in biomass may be due to different shoot densities or variability in leaf length and width (West and Larkum, 1983). The seasonal variation at Towra Point is similar to that found by Larkum *et al.* (1984) for *Zostera capricorni* measured over the same seasons in Botany Bay. Their result showed a 4-fold change in above ground biomass between summer and winter. The study found similar seasonal variability in Ukerabagh channel in Tweed Heads. Kirkman *et al.* (1982) also found biomass of *Z. capricorni* varying between summer and winter in Port Hacking, New South Wales. In south-eastern Australia Kerr and Strother (1990) found similar patterns for seasonal variability in *Z. muelleri*. The current experiment found the highest biomass of *Zostera capicorni* in Woolooware Bay (Towra Point) during spring followed by summer, autumn and the winter. The average mean biomass in Towra Point (Woolooware bay side) round the year is 78.0gm dry wt./m<sup>2</sup>.

Results from Ukeberagh Channel at Tweed Heads were somewhat different. The highest biomass was found during spring followed by winter and autumn, though on the whole biomass varied less than at Towra Point. The average mean biomass in Ukeberagh Channel is 208.68gm dry wt./m<sup>2</sup>, far higher than at Towra Point.

Both the sample sites were dominated by *Z. capicorni* but there was difference in shoot densities, length and width of the leaves, wave action, degree of pollution and other local conditions. The present study only examined the biomass difference between the 4 season of two locations. The biomass of *Zosteria capicorni* varies between sites depending on the morphological characteristics and local environmental conditions. West and Larkum (1979) compared five study sites and found large variations in biomass of *Posidonia australis*. According to West and Larkum (1983) biomass also vary depending on local conditions, such as depth, age of bed, degree of wave exposure and exposure to salinity changes. As seagrass meadows are known to contribute nutrients to estuarine food chains and to provide shelter for juvenile fish (Mann, 1972; Thayer *et al.*, 1975; Conacher et al.

1979; Middleton *et al.* 1984), the conservation of the beds of *Zostera capricorni* in both areas are of important.

Densities of biomass at Towra Point were consistently lower than in the Tweed Channel, a result which may be partly explicable by differing sediment regimes, in particular the movement of sediment into Woolooware Bay following erosion from Towra Point. Sedimentation in the seagrass beds was found to be in the order of 5cm in a 6-month period, nearly an order of magnitude higher than the adjacent mangrove. This high rate of sediment delivery is likely to create problems for the maintenance of biomass more than any latitudinal difference between the two seagrass sites.

Difference in biomass created important difference in the degree of replication required to provide an adequate sample. Within Ukerebagh Channel, biomass was consistently high, and the suggested level of replication (0.25mX 0.25m) was found to be sufficient to detect as significant a 10% variation. However, the situation at Towra Point was markedly different. Here, where biomass was low and variable, more than 15 quadrats would be required to detect as significant a change of 20%. The result highlights the need to determine the degree of replication for individual sites, rather than following suggestions as to the level of replication appropriate.

# **Chapter Four: sampling nekton in seagrass**

## 4.1 Introduction

Seagrasses are a valuable component of Australia's coastal resources for fisheries through food webs and the habitat for important stages in the life cycle. Seagrasses form a threedimensional canopy, which provides habitat for fish and crustaceans (Pollard, 1984). Seagrass meadows are the nursery grounds for many of Australia's commercial fish and crustacean species (Middleton et al. 1984; Bell and Pollard, 1989). In the Gulf of Carpentaria, along the east coast of Queensland and north-eastern Arnhem Land, juvenile tiger prawns and endeavor prawns use seagrass meadows as their nursery grounds (Staples et al. 1985; Poiner et al. 1987; Coles et al. 1993). The juveniles take shelter in seagrass bed for protection from predators. Minello and Zimmerman (1983) showed that the levels of predation on juvenile penaeids by fish were considerably lower in vegetated than in non-vegetated areas. An estuarine area that has been artificially modified and colonized by seagrass can provide a settlement habitat for large numbers of juvenile fish of economic value (McNeill et al., 1992). Seagrass also support complex food webs by virtue of their physical structure and primary production and are well known for their role as breeding grounds and nurseries for important crustacean, finfish and shell fish populations (Short and Coles, 2001).

Numerous methods have been used to sample fish populations associated with natural habitat as well as artificial structures, including scuba, controlled angling, electrofishing, trap nets, drop nets, seine net and rotenone. None of these methods can consistently provide accurate estimates of the entire fish assemblage associated with artificial structures because of inherent biases and limitations (Larson *et al.*, 1986). The limitations of most methods have not been evaluated and would be difficult to determine.

The composition of fish assemblages at any one location depends on the method of collection. Assemblages collected from the same meadow using two different methods (poisoning and trawling) can be more different than assemblages collected from different

meadows with the same method (Gray and Bell, 1986). Even the catches from different types of beam trawl from within one seagrass meadow vary in number of species and number of individuals (McNeill and Bell 1992).

In present study buoyant pop nets (Figure 4.1) were used to collect fish sample from the seagrass meadow at Woolooware Bay. The buoyant pop net which was used for collecting fish sample from the vegetated fish habitats by Larson *et al.* (1986), Michael *et al.* (1989), Connolly (1994) and Mazumder *et al.* (2004) has the advantage of providing instantaneous assessment of fish density in shallow water vegetated habitats.

The aim of the study is to analyse the seasonal variability of fish between spring and neap high tides, and to compare fish use of seagrass with adjoining mangrove and salt marshes. Further, an attempt has been made to test the efficiency of the buoyant pop net.

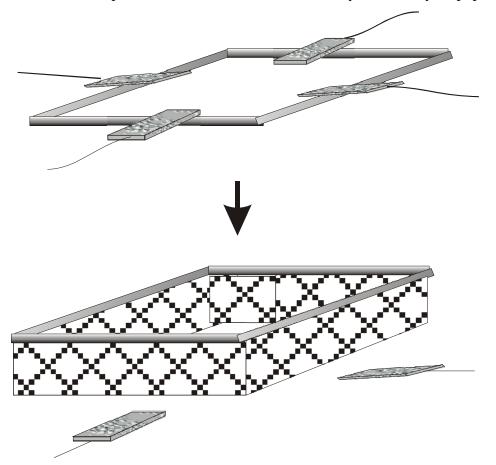


Figure 4.1 Configuration and operation of the buoyant pop net (after Mazumder 2004).

## 4.2. Study site and methods

## 4.2.1 Study Location

The study was conducted in seagrass meadow at Woolooware Bay which is one of the diverse tidal and estuarine wetlands in Sydney region of NSW (Figure 4.2). The seagrass meadow is comprised of *Zostera capricorni* (Larkum *et al.* 1984).

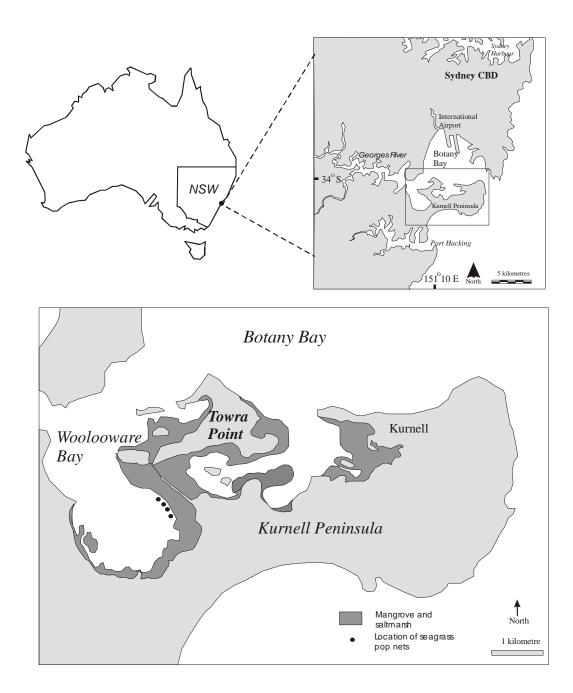


Figure 4.2 Location of pop nets within Woolooware Bay

#### 4.2.2 Fish collection

Buoyant pop nets (Michel *et al.* 1989, Connolly, 1994) were used in seagrass to sample fish during spring and neap tide. The pop nets consist of 2.0 mm nylon mesh and are 5.5 m by 5.5 m square, elevating to a maximum height of 2.5 meters. The top of the mesh was attached to 15 mm diameter PVC pipe, sealed for flotation. The base of the net is secured tightly to the marsh surface using long tent pegs, with the nets hidden prior to release inside shallow trenches. Nets were weighed down with four concrete blocks attached with nylon rope to a remote point 150 meters from the nets. The nets were released from this remote point at the of spring and neap tide. Four replicate pop nets were used in seagrass bed, placed randomly. Fish were retrieved using a sweeping hand net. Fish sample were collected monthly during spring high tides (>1.8m) and neap tides (< 1.2 m) from January 2002 to December 2002.

#### Net efficiency

A component of this study was to test the efficiency of the pop nets in seagrass. A seine net was used to capture fish exiting the saltmarsh. Collected fish species were then placed into the floating pop nets that had been released at spring tide. Care was taken to remove fish from the vicinity of the pop nets prior to net release. A total of 286 fish representing four different species were captured in preparation for this experiment. These were divided between two buoyant pop nets, raised at the peak of a spring tide. Fish were collected during the receding tide.

#### 4.2.3. Statistical methods

Differences in assemblages of fish between spring and neap high tide were examined using Bray-Curtis measures of dissimilarity (Bray and Curtis, 1957) with square root transformed data. Nonmetric multidimensional scaling (nMDS) plots were also used to demonstrate the patterns of fish assemblages in different seasons between high and neap tides (Clarke and Warrick 2001). Multivariate techniques such as ANOSIM (Clarke and Warrick 2001) was used to test the statistical significance of differences between the fish assemblages at spring and neap high tides. The contribution made by particular species to differences in species assemblages was determined using SIMPER (similarity percentages-species contribution, Clarke and Warrick 2001) for spring and neap high tide.

Univariate analysis (ANOVA) and post-hoc Student-Newmans-Kuels (SNK) tests were performed using GMAV5 (Underwood and Chapman, 1989) to determine the significance of differences of fish abundance between spring and neap tides in different seasons. Two way analysis of variance was used (Underwood, 1997) where season was considered as the first factor with four levels (1=Winter, 2=Spring, 3=Summer and 4=Autumn) which were orthogonal and fixed. Tide was considered as second factor with two levels (1=spring tide and 2= neap tide), which were orthogonal and fixed.

## 4.3 Results

## 4.3.1 Seasonal variability in fish and crustaceans abundance

A total of 3972 fish and crustaceans of eleven species were caught in both spring and neap tides in Woolooware Bay, Towra Point, using pop nets (Table 4.1) where *Zostera capricorni* was the main seagrass species. Eleven species were found and most of them were juveniles. Among all the species *Metapenaeus macleayi* was most abundant contributing 20.80% of the total catch, followed by *Macrobrachium intermedium* (16.01%), *Alpheus edwardsi* (15.21%), and *Pelates quadrilineatus* (11.78%). *Liza argenta* and *Acanthopagrus australis* are the least abundant in seagrass bed contributing 3.40% and 2.90% of total catch.respectively.

*P. quadrilineatus* was most abundant in high tide during spring (13.83%) and summer (13.25%), almost equal abundant in winter (11.97%) and low in high tide during autumn (10.53%). *Arenigobius bifrenatus* was most abundant in high tide during winter (8.61%), spring (7.01%) and summer (9.88%) but less abundant in high tide during autumn (5.76%). *Pseudogobius olorum* was most abundant in neap tide during winter (7.38%), summer (7.58%), autumn (9.52%) and less abundant in neap tide during spring (8.67%). *Ambassis jacksoniensis* was most abundant in neap tide at all season (8.38% in winter, 11.90% in spring, 10.17% in summer and 7.00% in autumn).

The crustacean species *M. macleayi* was most abundant in spring, summer and autumn in neap tide contributing 22.18%, 21.99%, and 21.36% respectively of the catch by number. *M. intermedium* was most abundant in high tide in winter (18.91%), summer (19.04%) and autumn (16.04%). *A. edwardsi* was highly abundant in high tide during winter (19.12%) and autumn (17.29%) and less abundant during summer (13.73%) in high tide.

Three crustacean (*M. macleayi, M. intermedium, A. edwardsi*) and three species of fish (*P. quadrilineatus, P. olorum* and *A. bifrenatus*) are common in seagrass bed as they are found almost all time. Fish abundance vary during seasons. Among 11 species found in seagrass bed four were of commercial importance (Table 4.1).

Table 4.1. Species caught from seagrass meadow with pop nets at Woolaware Bay, Towra Point from January 2002 – December 2002. The asterisk (\*) designates species of commercial and or recreational significance.

Family	Genus/species	Common name	% Contribution to catch									
			Winter	r	Spring	5	Summ	er	Autum	1	All sease	on
			(June-	Aug.)	(SepN	Nov.)	(Decl	Feb.)	(MarN	/Iay)		
			High	Neap	High	Neap	High	Neap	High	Neap	High	Neap
Gobiie	Arenigobius bifreentus	Bridled goby	8.61	6.39	7.01	6.05	9.88	8.87	5.76	7.18	7.79	7.16
Gobiidae	Pseudogobius olorum	Blue spot goby	6.72	7.38	9.81	8.67	5.54	7.58	8.77	9.52	7.84	8.31
Teraponidae	Pelates quadrilineatus	Trumpeter *	11.97	11.98	13.83	7.26	13.25	9.98	10.53	13.47	12.52	10.74
Tetraodontidae	Tetractenos hamiltoni	Common toad	2.73	2.20	4.90	2.42	4.82	3.33	4.51	3.59	4.25	2.91
Mugilidae	Liza argenta	Flat tail mullet *	0.64	4.79	2.63	4.64	3.86	2.40	3.52	4.85	2.58	4.15
Sparidae	Acanthopagrus australis	Yellow fin bream *	0.63	3.39	1.93	5.44	2.89	3.14	2.26	3.41	1.88	3.82
Ophichthidae	Ophisurus serpens	Eel fish	2.73	1.20	2.10	2.62	3.13	1.85	5.01	2.69	3.12	2.10
Chandidae	Ambassis jacksoniensis	Glass fish	5.67	8.38	9.81	11.90	6.99	10.17	5.51	7.00	7.20	9.31
Crustaceans												
Penaeidae	Metapenaeus macleayi	School prawn *	22.27	21.36	19.61	22.18	16.87	21.99	20.80	21.36	19.94	21.72
Atyidae	Macrobrachium intermedium	Grass shrimp	18.91	17.36	15.06	15.52	19.04	14.05	16.04	13.82	17.14	15.13
Alpheidae	Alpheus edwardsi	Common pistol shrimp	19.12	15.57	13.31	13.30	13.73	16.64	17.29	13.11	15.74	14.65

## 4.3.2 Fish abundance and species richness

## Spring high tide

ANOVA results showed that fish abundance in seagrass varied significantly among the seasons in spring high tide (P<0.01: Table 4.2).

Table 4.2: ANOVA results for fish abundance in spring high tide between seasons in seagrass using pop nets at Woolooware Bay, NSW: Cochran's Test C=0.3167

Source of variation	DF	MS	F	Р
Season	3	48.446	3.77	0.0107
Residual	524	12.855		
Total	527			

## Neap high tide

ANOVA results showed that fish abundance in seagrass did not vary among the seasons in neap tide (P>0.6515 : Table 4.3).

Table 4.3: ANOVA results for fish abundance in neap tide between seasons in seagrass using pop nets at Woolooware Bay, NSW: Cochran's Test C=0.2767

Source of variation	DF	MS	F	Р
Season	3	7.8737	0.55	0.6515
Residual	524	14.4387		
Total	527			

## Comparison of spring and neap high tide

ANOVA results showed that fish abundance in spring and neap tide vary between seasons (P<0.0039) and no significant difference was found in fish abundance between the tides (P>0.4389, Table 4.4).

Table 4.4: Two way analysis results for fish abundance between seasons and tides at Woolooware Bay, NSW: Cochran's Test C=0.1492

Source of variation	DF	MS	F	Р
Season	3	61.1550	4.48	0.0039
Tide	1	8.5464	0.63	0.4289
Season X Tide	3	10.0464	0.74	0.5305
Residual	1048	13.64783		
Total	1055			

The post-hoc SNK test for fish abundance between high and neap tide in different seasons showed fish abundance in winter in neap tide to be significantly higher than spring tide. Fish abundance between spring and neap tide in spring, summer and autumn did not vary significantly (Figure 4.3).

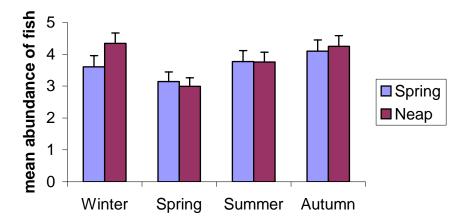


Figure 4.3: Mean abundance (+ SE) of fish in seagrass bed at Towra Point (Wooloware Bay) in different seasons using pop nets, n=132.

## Fish assemblages in spring and neap high tides

Non-metric multidimensional scaling (nMDS) ordinations between the tides showed different assemblages of fish in seagrass meadow (Figure 4.4).

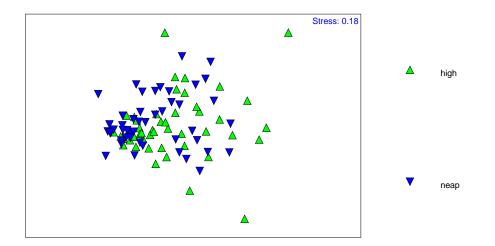


Figure 4.4. NMDS ordinations showing assemblages of fish in spring high and neap high tides at Woolooware Bay using pop nets.

Analysis of similarities (ANOSIM) results show that fish assemblages differed significantly between the spring and neap high tides (ANOSIM: P < 0.04). The dissimilarity between the tides appears primarily due to the higher abundance of *Ambassis jacksoniensis*, *Acanthopagrus australis*, *Metapenaeus macleayi*, and *Liza argenta* in the neap tide (Table 4.5).

Table 4.5: Contribution of species of dissimilarity between spring and neap high tides fish assemblages in the Woolooware Bay seagrass.

Group	spring Grou	p neap					
Species	Av.Abund	Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum.%	
Pistol shrimp	6.10	6.48	6.25	1.36	15.11	15.11	
School prawn	7.73	9.48	6.02	1.39	14.56	29.66	
Grass shrimp	6.65	6.60	5.64	1.35	13.64	43.30	
Glassfish	2.79	4.06	4.32	1.30	10.45	53.76	
Trumpeter	4.85	4.69	4.29	1.37	10.38	64.13	
Blue spot goby	3.08	3.62	3.86	1.37	9.32	73.46	
Bridal goby	3.02	3.12	3.45	1.27	8.35	81.80	
Flat tail mullet	1.00	1.81	2.19	1.13	5.31	87.11	
Yellow fin bream	0.73	1.67	2.10	1.03	5.09	92.20	

## **4.3.3 Results from efficiency experiment**

A total of 286 fish were released in to two pop nets but only 78 (27.3%) were retrieved from the two nets (Table 4.6). In net 1, there were 3 *Liza argenta*, 4 *Tetractenos hamiltoni*, 8 *Acanthopagrus australis* and 117 *Ambassis jacksoniensis* were released but retrieved 3 *Acanthopagrus australis*, 23 *Ambassis jacksoniensis*, all *Tetractenos hamilton* were retrieved but none of *Liza argenta* were retrieved. The retrieval rate in net 1 was 22.7%.

In net 2, there were 4 *Liza argenta*, 4 *Tetractenos hamiltoni*, 8 *Acanthopagrus australis*, and 138 *Ambassis jacksoniensis* released but retrieved 3 *Tetractenos hamiltoni*, 2 *Acanthopagrus australis*, and 43 *Ambassis jacksoniensis*. None of *Liza argenta* were retrieved. The retrieval rate in pop net 2 was 31.2%.

Table 4.6: Released and retrieved fish pop	ulation from two	buoyant pop	nets as a test of
net efficiency			

Species	Common name	Ne	et 1	Net 2	
		Released	Retrieved	Released	Retrieved
Liza argenta	Flat tail mullet	3	0	4	0
Tetractenos hamiltoni	Common toad	4	4	4	3
Acanthopagrus australis	Yellow fin bream	8	3	8	2
Ambassis jacksoniensis	Glass fish	117	23	138	43

## 4.3.4 Fish assemblages among the habitats (seagrass, mangrove and saltmarsh)

The experiments were conducted in tandem with an identical experiment performed by Debashish Mazumder in adjacent mangrove and saltmarsh (Mazumder, 2004). His data have been included with permission in an nMDS plot (Figure 4.5), which compares similarity of assemblages between adjacent seagrass, mangrove and saltmarsh habitat for five months between October, 2001 and February, 2002.

Non-metric multidimensional scaling (nMDS) ordinations showed different assemblages of fish in seagrass, saltmarsh and mangrove (Fig. 4.5). Analysis of similarities (ANOSIM) results show that fish assemblages differed significantly between the habitats (ANOSIM: P<0.0001).

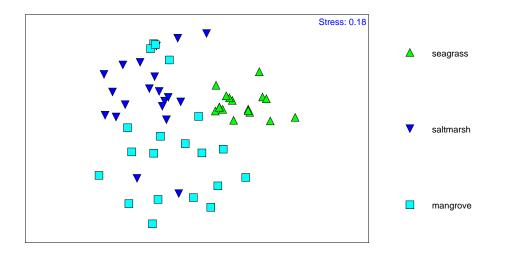


Figure 4.5. nMDS ordinations showing assemblages of fish in seagrass, mangrove and sltmarsh at Woolooware Bay using pop nets (including data from Mazumder, 2004).

## 4.4 Discussion

The results of the capture efficiency of pop nets demonstrated inter-specific difference in recapture rate. Low recapture of *Liza argenta* (zero recapture), high recapture rate for *Tetractenos hamiltoni* and moderate recapture rate for *Acanthopagrus australis* and *Ambassis jacksoniensis* were demonstrated for two released buoyant pop nets. Such results would under-represent some species particularly those able to jump out of the net (eg. *Liza argenta*). However the two nets were relatively consistent in their efficiency and comparison between nets and the environment remain valid. Mazumder (2004) found the average size of fish retrieved from the pop nets was consistently larger than from fyke nets and smaller fish caught in the pop net were not retrieved. He also suggested pop net catches were biased toward the larger size classes of fish. Further studies utilizing buoyant pop nets may wish to extend this efficiency test.

Although there were no significant differences in fish species abundance between the tides there were differences found between seasons. Almost all the species are present in both spring and neap high tides. Most of the fish species captured from *Zostera capricorni* bed in Woolooware Bay were juveniles. Settlement and recruitment of juvenile fish was estimated by Upston and Booth (2003), in natural and artificial seagrass in Botany Bay, NSW but there was no significant difference in abundance of recruits among the habitats. Middleton *et al.* (1984) found smaller fish species dominate *Zostera* meadow and larger fish species dominate the *Posidonia* site in Botany Bay, a finding reflected in this study. The study suggests that the *Zostera capricorni* meadow of Woolooware Bay is dominated by juvenile fish.

Fish abundances in Woolooware Bay were generally highest in autumn (Figure 4.3). State Pollution Control Commission (1981) reported the highest abundance of fish in autumn in Botany Bay. This seasonal peak in abundance was probably due to a peak in the seasonal recruitment of juveniles, as summer (February) is the peak in the abundance of fish eggs in the plankton and autumn is the peak in abundance of post larvae and small juveniles of commercial species (SPCC 1981).

Fish found in lower abundance in seagrass during the spring tide were those which Mazumder et al. (2004) found to be common in mangrove and saltmarsh during the same spring tides (*Alpheus edwardsi, Metapanaeus macleayi, Ambassis jacksoniensis, Pseudogobius olorum, Liza argeta, Acanthapagrus australis*). *Pelates quadrilineatus* was found not to differ between spring and neap tides in seagrass, and this species appears to be a seagrass specialist, rarely venturing into the mangrove or saltmarsh (Mazumder et al 2004).

A different assemblage of fish is found in spring high tide compare to neap tide. The monthly spring tide had the lower diversity and abundance of fish species than the neap tide. These are species which were found to moving into the adjacent mangrove and salt marsh by Mazumder *et al* (2004). Indeed the nMDS plot (Figure 4.5) shows clear

difference in the fish communities in the three adjacent habitats at the peak of the spring tide, with seagrass exhibiting lower concentration of fish species.

The results strongly suggest nekton move out of the seagrasses during the spring tides and into the adjacent inter tidal habitats, and less so during the neap tide when only seagrass and mangrove are covered. Nekton using estuarine habitats are highly mobile and move readily between multiple habitat types regularly over a tidal cycle or during the course of their life cycle (Skilleter and Loneragan, 2003). Many species move into intertidal areas during the flood tide and retreat to the shallow subtidal during the ebb flow (Rozas and Odum 1987; Hettler 1989; Kneib and Wagner 1994; Thomas and Connolly 2001). This illustrates the importance of viewing estuarine wetlands as a mosaic of inter-connected habitats.

# **Chapter Five: Discussion and Recommendations**

#### **5.1 Important results**

Commonly used protocols for biological monitoring are seldom subject to rigorous assessment. More often, techniques used are justified by simple reference to authority which can become an expedient means to propagate unreliable methods or unreplicable results (Peters 2001). This thesis sought to examine and apply seagrass monitoring protocols with a view to analyzing their efficiency and reliability. Primarily, it was hoped that the outcomes of the thesis might lead to recommendations concerning optimal sampling strategies for *Zostera capricorni*. In the process, some new insights were gained into the functioning of *Zostera* as a fish habitat.

The ARCView Geographic Information System was used to construct vegetation polygons of seagrass distribution on the Tweed River, and in the Ukerebagh Channel annually over a 5-year period. There were a number of reasons for choosing the Tweed River. Firstly, the quality of the aerial photograph coverage was excellent. The clarity of the water over the study period allowed for relatively easy identification of the seagrass boundaries. This eliminated identification errors from the analysis, allowing this work to concentrate on errors relating to boundary definition. The Tweed River was also subject to major entrance modification works over the period of study. The opportunity existed to study a system in flux, as increased tidal velocities modified tidal shoals, potentially redistributing seagrass.

The mapping showed relatively low inherent error in boundary definition. A 2.9% discrepancy was found when the Ukerebagh Channel was mapped twice from the year 2000 photographs. On this basis, the inter-annual variability was deduced to be real, being a consistent increase in extent in the range of 5-12% per annum. There was no evidence that this increase was the result of the Tweed sand bypassing, which commenced in 2001, as increases in extent were occurring before this time. More likely, the increases in seagrass extent within the Ukerebagh Channel related to the *El Nino* drought, with natural extention unimpeded by flood scouring or sedimentation.

Importantly, the study suggests inter-annual variability relevant to any monitoring study. This can be contrasted to the mapping of mangrove and saltmarsh, in which inter-annual variability is insignificant in most cases (Saintilan and Wilton 2000). One clear recommendation is that seagrass extent could usefully be mapped more frequently than the 5-yearly period recommended by Saintilan and Wilton (2000) for mangrove and saltmarsh.

Estimates of biomass were derived from standard replicate 0.25m x 0.25m quadrats following Larkum et al. (1984), Larkum and West (1990), Kirkman and Cook (1982), Mellors (1991) and Lanyon and Marsh (1995). The experiment contrasted two sites of similar geomorphic setting. Ukerebagh Channel on the Tweed River, and Woolooware Bay with Botany Bay are both shallow, sandy marine deltaic settings supporting stands of *Zostera capricorni*. Significant differences were found in the degree of replication required to identify significant changes in seagrass biomass at the two sites. Ukerebagh Channel supported relatively dense stands of *Z. capricorni* with low intra-site variability. Here 8 replicates were sufficient to detect 10 percent change at P = 0.05. Towra Point presented a contrast, in which 15 replicates were required to detect a similar level of change. Towra Point has suffered from increased sedimentation relating to alterations in current velocities at Towra Point, and the result highlights the greater degree of replication required to determine significance changes in disturbed systems.

Previous research on estuarine habitats has led to significant advances in our understanding of the importance of specific features of habitats, such as the complexity of mangrove root systems on finfish and crustacean abundance. However, this approach has treated different types of habitat as isolated and separate patches rather than as a 'mosaic' of interlinked habitats. Recent studies have shown that fish species utilise habitat mosaics and move between multiple habitat types regularly over a tidal cycle or during the course of their life cycle (Skilleter and Loneragan, 2003). Fluxes in the density of fish and the availability of food between the interlinked habitats of seagrass and mangrove, and the contribution of this mosaic to the fishery is worthy of quantification.

The fish populations in the seagrass at Towra Point were sampled using buoyant pop nets, concurrently with surveys using the same technique in the mangrove and saltmarsh performed by Mazumder (2004). Fish communities differed significantly from those sampled in adjacent mangrove and saltmarsh. Differences in fish assemblages between spring high tides, neap high tides and low tides are attributed to movements of fish between seagrass and adjacent mangrove and saltmarsh. This mosaic of habitats is utilized by a number of species over a tidal cycle, with seagrass providing an important low-tide refuge for many species utilizing mangrove and saltmarsh at high tide.

The result is of significance for a number of reasons. Firstly, it underscores the importance of managing estuaries with consideration of linkages between habitats. The value of seagrass, mangrove and saltmarsh might differ depending on their placement with respect to adjacent habitats. It may be that the finding of Mazumder et al (2004) that the Towra Point mangrove and saltmarsh fish assemblage is more diverse than Homebush Bay and Allens Creek can be attributed to the absence of seagrass adjacent to these locations, rather than the marine effects cited by the authors. Future studies would do well to consider the influence of seagrass on the diversity and abundance of fish assemblages in mangrove and saltmarsh in a more controlled experimental design.

Limitations in the efficiency of buoyant pop nets were exposed in a novel experiment which demonstrated differences in escape rates between species. Flat-tailed mullet (*Liza argenta*) are likely to be under-represented in experiments using this technique, as are bream (*Acanthropagrus australis*). This is an important consideration when sampling in these environments. The buoyant pop net has considerable advantages over fyke nets and seine nets in a seagrass setting, because catch can be translated directly into density. However, this is the first study that has considered catch efficiency in anything but a comparative study. Clearly, the pop net will require some design modifications if they are to be employed in inter-site comparisons of fish assemblages.

## 5.2 Recommendations regarding sampling

A number of recommendations can be made regarding optimal sampling protocols for monitoring seagrass in the region. These can be summarized as follows:

- 1. Seagrass should be mapped more frequently than mangrove and saltmarsh if changes in distribution are to be detected in time series. The 5-year period nominated for mangrove and saltmarsh is inadequate as applied to seagrass.
- 2. The degree of replication required for biomass assessment varies depending on the characteristics of the seagrass community. Patchy disturbed seagrass communities require 15-20 replicate 0.25m by 0.25m quadrats to detect 20% change at P<0.05. Stable, dense seagrass communities require a lesser degree of replication, and eight replicates may be sufficient to identify even 10% change at P<0.05. In all cases power analysis should be performed to determine whether an adequate sample has been attained.
- 3. The buoyant pop net should be modified to more adequately retain fish caught within the net at the time of release. This might best be done by securing an additional barrier to escape after release, such as a heightened wall. The net could, for instance, be propped up to an extended height above the water level with the use of vertical stakes. This would prevent the escape of fish over the top of the buoyant rim, which was clearly happening in the case of *Liza argenta* (which can jump), and most probably *Acanthropagrus australis*.

## 5.3 Recommendations for future research

The study has suggested a number of lines of enquiry which might further clarify the ecological function of seagrass and the means of studying these interactions. While this study has identified one source of mapping error (boundary definition in identical images), other sources of mapping error could be studied experimentally. One example would be the comparison of results where seagrass boundaries were not so clearly delinieated. This might occur where image quality is poor, (either due to poor photograph quality or poor water quality), or where seagrass grades into fluvial muds, rather than the quartz sand found on the Tweed.River It would be useful to gain an appreciation of circumstances in which seagrass mapping using on-screen digitizing from colour images is inappropriate, and whether the infra-red specra might provide better results.

The study has suggested important linkages between seagrass and adjacent wetland habitats over the course of the diurnal and neap-spring tidal cycles. The importance of habitat mosaics and linkages cannot be overstated, and this new area of research deserves more thorough treatment. As previously suggested, a controlled study might be envisaged which compares mangrove and saltmarsh utilization by fish in the presence and absence of seagrass. Such an experiment could either use natural habitat mosaics or incorporate a BACI design (Underwood 1991, 1992) through the removal or planting of seagrass.

The study has showed the abundance of commercial and ecological important fish species in the seagrass during high and neap tidal cycles. To what extent seagrass, seagrass epiphytes and epibenthic microalgae provide food for these species were beyond the scope of the current research. However, a detailed assessment of nutritional contribution of seagrass, epiphyte and epibenthic microalgae to the commercial and ecological fish species is therefore desirable.

## **REFERENCES:**

Abal, E.G., Loneragan, N., Bowen, P., Perry, C.J., Udy, J.W. and Dennison, W.C. (1994) Physiological and morphological responses of the seagrass *Zostera capicorni* Aschers. to light intensity. *Journal of Experimental Marine Biology and Ecology* 178: 113-129.

Bell, J.D. and Pollard, D.A. (1989) Ecology of fish assemblages and fisheries associated with seagrasses. In: A.W.D. Larkum, A.J. McComb and S.A. Shephard (Eds.), *The Biology of Seagrasses – An Australian Perspective*. Elsevier, Amsterdam, pp. 565-609.

Blaber, S.J.M., Brewer, D.T., Salini, J.P., Kerr, J.D. and Conacher, C., (1992) Species composition and biomasses of fishes in tropical seagrasses at Groote Eylandt, Northern Australia. *Estuarine Coastal and Shelf Science* 35: 605-620.

Bray, J.R. and Curtis, J.T. (1957) An ordination of the upland forest communities southern Wisconsin. *Ecological Monographs* 27: 325-349.

Butler, A. and Jerakoff, P. (1999) *Seagrass in Australia: Strategic review and development of an R & D plan*, CSIRO Publishing, Melbourne.

Charpy-Roubaud, C. and Sournia, A. (1990) The comparative estimation of phytoplanktonic and microphytobenthic production in the oceans. *Marine Microbial Food Webs* 4: 31-57.

Clarke, K.R., and Warwick, R.M. (2001) *Changes in marine communities: An approach to statistical analysis and interpretation*, 2<sup>nd</sup> edition. PRIMER-E; Plymouth, United Kingdom.

Coles, R.G., Lee Long, W.J., Watson, R.A. and Derbyshire, K.J. (1993) Distribution of seagrasses and their fish and penaeid prawn communities in Cairns Harbour, a tropical estuary, Northern Queensland. *Australia. Australian Journal of Marine and Freshwater Research* 44: 193-201.

Conacher, M.J., Lansing, W.J.R. and Larkum, A.W.D. (1979) Ecology of Botany Bay. II. Aspects of the feeding ecology of the fanbellied leatherjacket, *Monocanthus chinensis* in *Posidonia australis* seagrass beds in Quibray Bay, Botany Bay, New South Wales. *Australian Journal of Marine and Freshwater Research* 30: 387-400.

Connolly, R.M. (1994) Comparison of fish catches from a buoyant pop net and a beach seine net in a shallow seagrass habitat. *Marine Ecology Progress Series* 109: 305-309.

Costanza, R., d'Arge, R., Sutton, P. and van der Belt, M. (1997) The value of the world's ecosystem services and natural capital. *Nature* 387: 253-260.

CSIRO (1997) The facts, CSIRO Marine Research, Understanding our Oceans.

Day, J.W., Hall, C.A.S., Kemp, M.W. and Yanez-Arancibia, A. (1989) *Estuarine Ecology*. John Wiley and Sons. pp 558.

Downing, J. A. and Anderson, M. R. (1985) Estimating the standing biomass of aquatic macrophytes. *Canadian Journal of Fisheries and Aquatic Science* 42: 1860-1869.

Duarte, C.M. and Cebrian, J. (1996) The fate of marine autotrophic production. *Limnol.Oceanogr.* 41: 1758-1766.

Dyson, A., Victory, S. and Connor, T. (2001) Sand bypassing the Tweed River Entrance: *An Overview Paper presented in Coasts and Ports 2001*, Conference. I.E. Aust., Gold Coast, Queensland. Environment Australia (2000) Australia *State of the Environment Report 2001* - Coasts and Oceans.

Gambi, M.C., Nowell, A.R.M. and Jumars, P.A. (1990) Flume observations on flow dynamics in *Zostera marina* (eelgrass) beds. *Marine Ecology Progress Series*, 61: 159-169.

Gray, C.A. and Bell, J.D. (1986) Consequences of two common techniques for sampling vagile macrofauna associated with the seagrass *Zostera capricorni*. *Marine Ecological Progress Series* 28: 43-48.

Gray, C.A., McElligot, D.J. and Chick, R.C. (1996) Intra-and inter-estuary differences in assemblages of fishes associated with shallow seagrass and bare sand. *Australian Journal of Marine and Freshwater Research* 47: 723-735.

Green, E., Clark, C. and Edwards, A. (2000b) Image classification and habitat mapping. *Remote Sensing Handbook for Tropical Coastal Management* (ed. Alasdair, J. Edwards). UNESCO Publishing, France, pp 316.

Grey, W.F. and Moffler, M.D. (1978) Flowering of the seagrass *Thalassia testudinum* (Hydrocharitaceae) in the Tampa Bay, Florida area. *Aquatic Botany* 5: 251-259.

Harris, M., King, R.J. and Ellis, J. (1979) The eelgrass *Zostera capicorni* in Illawarra Lake New South Wales. Proc. Linn. Soc. NSW 104(1): 23-33.

Hartog, D. (1970) The Seagrasses of the World. North Holland, Amsterdam pp 275.

Hemminga M.A., and Duarte C.M. (2000) *Seagrass Ecology*. Cambridge University Press.

Hettler, W.F. (1989) Nekton use of regularly-flooded saltmarsh cordgrass habitat in North Carolina, USA. *Marine Ecological Progress* Series 56: 111-118.

Higginson, F.R. (1965) The distribution of submerged aquatic angiosperms in the Tuggerah Lakes system. Proc.Linn.Soc.NSW, 90: 328-334.

Hill, B.J. and Wassenberg, T.J. (1993) Why are some prawns found in seagrass? An experimental study of Brown (*Penaeus esculentus*) and Grooved (*P.semisulcatus*) Tiger prawns. *Australian Journal of Marine and Freshwater Research* 44: 221-227.

Hutomo, M., and Peristiwady, T. (1996) Diversity, abundance and diet of fish in the seagrass beds of Lombok Island, Indonesia. *In Seagrass Biology: Proceedings of an International Workshop* (eds. John Kuo, Ronald C. Phillips, Diana I. Walker and Hugh Kirkman). University of Western Australia, Perth. pp 385.

Hyder Consulting Pty. Ltd., Patterson Britton Partners Pty. Ltd. and WBM Oceanics Australia joint venture. (1997) Tweed River Entrance Sand Bypassing Project, Permanent Bypassing System, NSW Department of Land and Water Conservation and Queensland Department of Environment and Heritage, Sydney and Brisbane.

Inglis, G.J. and Smith, M.L. (1995) An examination of observer bias as a source of error in surveys of seagrass shoots. *Australian Journal of Ecology* 20(2): 173-281.

Kendrick, G.A., Eckersley, J. and Walker, D.I. (1999) Landscape-scale changes in seagrass distribution over time: a case study from Success Bank, Western Australia. *Aquatic Botany* 65: 293-309.

Kendrick, G.A., Hegge, B.J., Wyllie, A., Davidson, A. and Lord, D.A. (2000) Changes in seagrass cover on Success and Parmelia Banks, Western Australia between 1965 and 1995. *Estuarine, Coastal and Shelf Science* 50: 341-353.

Kennish, M.J. (1986) *Ecology of Estuaries*, Volume II: Biological Aspects. CRC Press, pp 391.

Kerr, E.A and Strother, S. (1990) Seasonal changes in standing crop of *Zostera muelleri* in south-eastern Australia. *Aquatic Botany* 38: 369-376.

Kikuchi, T. (1980) Faunal relationship in temperate seagrass beds. *In Handbook of Seagrass Biology: an Ecosystem Perspective* (eds. R.C. Phillips and C.P. McRoy), Garland STPM Press, New York, pp 353.

King, R.J. (1981) Marine angiosperms: *seagrasses. In Marine Botany*: An Australian Perspective (eds. N. Clayton and R. J. King), Longman Cheshire, pp 468.

Kirkman, H. (1997) Seagrasses of Australia: State of the Environment Technical Paper Series (Estuaries and the Sea), Department of Environment, Canbera pp 36.

Kirkman, H. (1996) Baseline and monitoring methods for seagrass meadows. *Journal of Environmental Management* 47: 191-201.

Kirkman, H. and Cook, I.H. (1982) Biomass and growth of *Zostera capricorni* Aschers. in Port Hacking, NSW, Australia. *Aquatic Botany* 12: 57-67.

Kirkman, H., and Reid, D.D. (1979) A study of the role of the seagrass *Posidonia australis* in the carbon budget of an estuary. *Aquatic Botany* 7: 173-183.

Kirkman, H. (1978) Decline of seagrass in northern areas of Moreton Bay, Queensland. *Aquatic Botany* 5: 63-76.

Kneib, R.T. and Wagner, S.L. (1994) Nekton use of vegetated marsh habitats at different stages of tidal inundation. *Marine Ecological Progress Series* 106: 227-238.

Kuo, J. and McComb, A.J. (1989) Seagrass taxonomy, structure and development. In *Seagrasses: A Treatise on the Biology of Seagrasses with Special Reference to the Australian Region*. A.W.D. Larkum, A.J. McComb and S.A. Shepherd (eds), Elsevier, North Holland, pp 6-67.

Lanyon, J.M., and Marsh, H. (1995) Temporal changes in the abundance of some tropical intertidal seagrasses in North Queensland. Aquatic Botany 49: 217-237.

Larkum A.W.D. and West R.J. (1990) Long-term changes of seagrass meadows in Botany Bay, Australia. *Aquatic Botany* 37: 55-70.

Larkum, A.W.D., McComb, A.J. and Shepherd, S.A. (eds) (1989) Seagrasses: A treatise on the biology of seagrasses with special reference to the Australian region. Elsevier, North Holland, pp 1-5.

Larkum, A.W.D., Collett, L.C., and Williams, R.J., (1984) The standing stock, growth and shoot production of *Zostera capricorni* aschers. In Botany Bay, New South Wales, Australia. *Aquatic Botany* 19: 307-327.

Larkum, A.W.D., (1977) Recent research on seagrass communities in Australia. In: C.P. McRoy and C. Helfferich (Editors), *Seagrass Ecosystems, A scientific Perspective*. Marcel Dekker, New York, pp 247-262.

Larkum, A.W.D. (1976) Ecology of Botany Bay I. Growth of Posidonia australis (Brown) Hook. f. in Botany Bay of the Sydney Basin. *Australian Journal of Marine and Freshwater Research* 27: 117-127.

Larson, E.W., Johnson, D.L., and Lynch, Jr. W.E. (1986) A buoyant pop net for accurately sampling fish at artificial habitat structures. *Transactions of the American Fisheries Society* 115: 351-355.

Long, B.G, Skewes, T.D. and Poiner, I.R. (1994) An efficient method for estimating seagrass biomass. *Aquatic Botany* 47: 277-291.

Mann, K.H., (1972) Macrophyte production and detritus food chains in coastal waters. *Mem. Inst. Ital. Idrobiol.*, 29: 353-383.

Marba, N. and Duarte, C.M. (1998) Rhizome elongation and seagrass clonal growth. *Marine Ecology Progress Series* 174: 269-280.

Mazumder, D. (2004) Contribution of saltmarsh to temperate estuarine fish in Southeast Australia. PhD Thesis, Faculty of Arts and Science, Australian Catholic University.

Mazumder, D., Saintilan, N. and Williams, R.J. (2004) Temporal variation in fish catch using pop nets in mangrove and saltmarsh flats at Towra Point, NSW, Australia. *Wetlands Ecology and Management* (accepted for publication).

McNeill, S. E. and Bell, J. D. (1992) Comparison of bean trawls for sampling macrofauna of *Posidonia* seagrass. *Estuaries* 15: 360-367.

McNeill, S. E., Worthington, D. G., Ferrell, D. J. and Bell, J. D. (1992) Consistently outstanding recruitment of five species of fish to a seagrass bed in Botany Bay, NSW. *Australian Journal of Ecology* 17: 359-365.

McQuade, M., Jackson, L.A. and Corbett, B.B. (2001) Tweed River Entrance Sand Bypassing Pre commissioning Dredging Works, Technical paper.

Meehan, A. J. (2002) Conservation status of the seagrass *Posidonia australis* Hook f. in south east Australia, PhD Thesis, Faculty of science, University of Wollongong, pp 230.

Meehan, A. J. (1997) Historical changes in seagrass, saltmarsh and mangrove communities in Lake Merimbula and Lake Pambula. Honours thesis (unpub.), University of Wollongong, pp 108.

Mellors, J.E. (1991) An evaluation of a rapid visual technique for estimating seagrass biomass. *Aquatic Botany* 42: 67-73.

Michael, R.D., Holland-Bartels, L.E. and Zigler, S.J. (1989) Comparison of fish catches with buoyant pop nets and seines in vegetated and nonvegetated habitats. *North American Journal of fisheries Management* 9: 249-253.

Middleton, M.J., Bell, J.D., Burchmore, J.J., Pollard, D.A. and Pease, B.C. (1984) Structural differences in the fish communities of *Zostera capricorni* and *Posidonia australis* seagrass meadows in Botany Bay, New South Wales. *Aquatic Botany* 18: 89-109.

Minello, T.J., and Zimmerman, R.J. (1983) Fish predation on juvenile brown shrimp, Penaeus aztecus Ives: the effect of simulated Spartina structures on predation rates. *Journal of Experimental Marine Biology and Ecology* 72: 211-231.

Moriarty, D.J.W., Boon, P.I., Hansen, J.A., Hunt, W.G., Poiner, I.R., Pollard, P.C., Skyring, G.W. and White, D.C., (1984) Microbial biomass and productivity in seagrass beds. *Geomicrobiology Journal* 4: 21-51.

Mumby, P. and Green, E. (2000b) Mapping Seagrass Beds. *Remote Sensing Handbook for Tropical Coastal Management* (ed. Alasdair J. Edwards). UNESCO Publishing, France. pp 316.

Neverauskas, V.P. (1987) Monitoring seagrass beds around a sewage sludge outfall in South Australia. *Marine Pollution Bulletin* 18: 158-164.

NSW Fisheries (1999b) Seagrasses in New South Wales, NSW Fisheries, Sydney.

NSW Fisheries (2004) Fish Care – Saving our Seagrasses Fish note DF/29.

Poiner, I.R. and Peterken, C. (1995) Seagrasses. In The State of the Marine Environment Report for Australia Technical Annex 1: The Marine Environment (eds. Zann, L.P. and Kailola, P.), Great Barrier Reef Marine Park Authority, Townsville, pp 107-117.

Poiner, I.R., Staples, D.J., and Kenyon, R. (1987) Seagrass communities of the Gulf of Carpentaria, Australia. *Australian Journal of Marine and Freshwater Research* 38: 121-131.

Poiner, I.R., Walker, D.I. and Coles, R.G. (1989) Regional Studies – Seagrasses of Tropical Australia. *Biology of Seagrasses: a Treatise on the Biology of Seagrasses with Special Reference to the Australian Region* (eds. Larkum, A.W.D., McComb, A.J., and Shepherd, S.A.) Elsevier Amsterdam, pp 279-303.

Peters R.H. (2001) A Critique for Ecology. Cambridge University Press 366pp.

Pollard, D. A. (1984) A review of ecological studies on seagrass-fish communities with particular reference to recent studies in Australia. *Aquatic Botany* 18: 3-42.

Preen, A., Thompson, J. and Corkeron, P., (1992) Wildlife and Management: Dugongs, Waders and Dolphins. In: Moreton Bay in the Balance. O.N. Crimp (Editor), Australian Littoral Society, Brisbane, pp. 59-68.

Preen, A.R., Lee Long, W.J., and Coles, R.G. (1995) Flood and cyclone related loss, and partial recovery, of more than 1000km<sup>2</sup> of seagrass in Hervey Bay, Queensland, Australia. *Aquatic Botany* 52: 3-17.

Robertson, E.L. (1984) Seagrasses. Chapter 6 in *The Marine Benthic Flora of South Australia. Part 1.* Womersley, H.B.S. ed. Government Printer Adelaide.

Roblee, M.B. and Zieman, J.C. (1984) Diel variation in the fish fauna of a tropical seagrass feeding ground. *Bulletin of Marine Science* 34: 335-345.

Rogers, K., Saintilan, N., Hossain, M.K. and Wilson, N. (2003) Tweed River Estuary, Estuarine Vegetation Monitoring Program, NSW Department of Infrastructure Planning and Natural Resource and The Queensland Environmental Protection Agency.

Rotherham, D. (1999) Comparison of fish assemblages in *Zostera capricorni* and *Posidonia australis* seagrass beds in three estuaries of south eastern Australia. Honours Thesis (unpub.) University of Wollongong, pp 95.

Rozas, L.P. and Odum, W.E. (1987) The role of submerged aquatic vegetation in influencing the abundance of nekton on contiguous tidal fresh-water marshes. *Journal of Experimental Marine Biology Ecology* 114: 289-300.

Saintilan N. and Wilton K. (2000). Protocols for the mapping of mangrove and saltmarsh vegetation. Report prepared for the Department of Land and Water Conservation, Coastal Weltands Unit Technical Report 2000/01.

Seddon, S., Connolly, R.M., and Edyvane, K.S. (2000) Large scale dieback in northern Spencer Gulf, South Australia. *Aquatic Botany* 66: 297-310.

Short, F. T. and Coles, R. (eds) (2001) *Global Seagrass Research Methods*. Elsevier Publishers, The Netherlands pp 482.

Skilleter, A.G. and Loneragan, N.R. (2003) Assessing the importance of coastal habitats for fisheries, biodiversity and marine reserves: a new approach taking into account "habitat mosaics". *Proceedings of the World Congress on Aquatic Protected Areas*. Cairns, Australia; August 2002. In Press.

Smith, S.V. (1981) Marine macrophytes as a global carbon sink. Science 211: 838-840.

SoEAC (1996) State of the Environment Australa 1996, Estate of the Environment Advisory Council, Department of the Environment, Sport and Territories, CSIRO Publishing, Victoria.

Staples, D. J., Vance, D.J. and Heales, D.S. (1985) Habitat requirements of juvenile penaeid prawns and their relationship to offshore fisheries, in Second Australian national prawn seminar, eds. P.C. Rothlisberg, B.J.Hill and D.J. Staples, Cleveland, Queensland: pp 47-54.

State Pollution Control Commission (1981) The Ecology of Fish in Botany Bay, pp 1-78.

Steffensen, D.A. and McGregor, F.E. (1976) The application of aerial photography to Estuarine Ecology. *Aquatic Botany* 2: 3-11.

Thayer, G.W., Wolfe, D.A. and Williams, R.B. (1975) The impact of man on seagrass ecosystems. *Am.Sci.*, 63: 288-296.

Thomas, B.E. and Connolly, R.M. (2001) Fish use of subtropical saltmarshes in Queensland, Australia: relationship with vegetation, water depth and distance on to the marsh. *Marine Ecological Progress Series* 209: 275-288.

Underwood, A. J. (1991) Beyond BACI: Experimental designs for detecting human environmental impacts on temporal variations in natural populations. *Australian Journal of Marine and Freshwater Research*. 42: 569-87.

Underwood, A. J. (1992) Beyond BACI: the detection of environmental impacts on populations in the real, but variable, world. *Journal of Experimental Marine Biology and Ecology* 161: 145-178.

Underwood, A.J. (1997) Experiments in Ecology: Their Ecological Design and Interpretation Using Analysis of Variance. University Press, Cambridge, UK.

Underwood, A.J. and Chapman, M.G. (1989) GMAV5 for Windows, Institute of Marine Ecology, University of Sydney, Australia.

Upston, J. and Booth, D. J. (2003) Settlement and density of juvenile fish assemblages in natural, *Zostera capricorni* (Zosteraceae) and artificial seagrass beds. *Environmental Biology of Fishes* 66(1): 91-97.

Walker, D., Dennison, W. and Edgas, E. (1999) Status of Australian Seagrass Research and Knowledge, CSIRO Publishing.

Walker, D.I. (1989) Regional studies – seagrass in Shark Bay, the foundation of an ecosystem, in Biology of seagrasses; a treatise on the biology of seagrasses with special reference to the Australian region (eds. Larkum, A.W.D., McComb, A.J., and Shepherd, S.A.) Amsterdam, Elsevier pp 841.

Watford, F.A. and Williams, R.J. (1998) Inventory of Estuarine Vegetation in Botany Bay, With Special Reference to Change in the Distribution of Seagrass, Coast and Clean seas, Project No. 97/003741. NSW Fisheries Final Report Series No. 11 pp 49.

Watson, R.A., Coles, R.G. and Lee Long, W.L. (1993) Simulation estimates of annual yield and landed value for commercial Penaeid prawns from a tropical seagrass habitat, Northern Queensland, Australia. *Australian Journal of Marine and Freshwater research* 44: 211-219.

West, R.J. (1990) Depth-related structural and morphological variations in an Australian *P. australis* seagrass bed. *Aquatic Botany* 36: 153-166.

West, R.J., Larkum, A.W.D. and King, R.J. (1989) 'Regional studies-seagrasses of south eastern Australia', In Biology of Seagrasses: A treatise on the Biology of Seagrases with Special Reference to the Australian Region, (eds. A.W.D. Larkum, A.J McComb & S.A Shepherd). Elsevier, Amsterdam, pp 230-260.

West, R.J., Thorogood, C.A., Walford, T.R. and Williams, R.J., (1985) An Estuarine Inventory for New South Wales, Australia. Fisheries Bulletin No. 2, Department of Agriculture, NSW, Australia, pp 140.

West, R.J. (1983) The seagrasses of New South Wales estuaries and embayments. *Wetlands* 3(1): 34-44.

West, R.J. and Larkum, A.W.D. (1983) Seagrass primary production – a review. *Proc.Linn.Soc.* NSW, 106(3): 213-223.

West, R.J. and Larkum, A.W.D. (1979) Leaf productivity of the seagrass, *Posidonia australis* in eastern Australian waters. *Aquatic Botany* 7: 57-65.

Williams R. J. and Meehan A. J. (2001) Historical changes in distribution of estuarine macrophytes of Port Hacking, NSW, Australia. Final Report to Coast and Clean Seas.

Williams, R.J. and Watford, F. (1999) Inventory of estuarine vegetation in Botany Bay, with special reference to change in the distribution of seagrass. NSW Fisheries Final Report Series No. 11, pp 44.

Wolterding, M. (2000) The life history of *Posidonia australis* Hook.f. PhD thesis, Sydney University pp 411.

Wood, N., and Lavery, P. (2000) Monitoring seagrass ecosystem health: The role of perception in defining health and indicators. *Ecosystem Health* 6(2): 134-148.

Young, P.C., (1978) Moreton Bay, Queensland: a nursery area for juvenile penaeid prawns. *Australian Journal of Marine and Freshwater Research* 28: 745-773.

Zann, L.P. (1995) Our Sea, Our Future. Major findings of the state of the Marine Environment Report for Australia, Great Barrier Reef Marine Park Authority for DEST. Ocean Rescue 2000 program.